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Genetic evaluation of population structure in white marlin (Kajikia albida): the importance of statistical power

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Introduction

Population genetic studies of marine fishes provide information to fisheries managers useful for the identification of biological units relevant for assessment and management, and for maintaining unique genetic variation (Allendorf et al., 1987; Ward, 2000; Ovenden et al., 2015). Despite their utility, these studies are particularly challenging for many marine fishes because the large effective population sizes of these species limit genetic drift, resulting in low levels of genetic differentiation among populations (Ward et al., 1994; Waples, 1998). A high level of statistical power is required to detect genetic differentiation in population genetic studies of marine fishes, and can be achieved by evaluating larger numbers of molecular markers and/or samples per putative population (Waples, 1998; Ryman et al., 2006). Although the former strategy is facilitated by currently available laboratory methodologies, obtaining appropriate sample sizes for population genetic studies of relatively rare-event species can be difficult.

Additional considerations for population genetic studies of highly migratory marine fishes are associated with sampling designs appropriate for these species. Highly migratory marine fishes are capable of long distance movements, and opportunistic experimental designs based on the sampling of geographically distant locations may not be informative because individuals of the same stock could be sampled from multiple locations depending on the time of year (Graves et al., 1996; Carlsson et al., 2007; Graves and McDowell, 2015). Many highly migratory marine fishes also display seasonal mixed assemblages, and opportunistic sampling may lead to sample collections representative of more than one stock, resulting in a noisy genetic signal that may mask genetic differentiation (Waples, 1998; Bowen et al., 2005). For highly migratory marine fishes that display population structure,
populations are most likely to separate at the time of spawning (Graves et al., 1996; Carlsson et al., 2007; Graves and McDowell, 2015). Replacing opportunistic sampling with a biologically informed design that targets larvae and/or reproductively active adults improves the ability to detect population subdivision in these species, if it exists. This concept is illustrated in Atlantic bluefin tuna (Thunnus thynnus) for which eastern and western Atlantic stocks were initially recognized based on the presence of distinct spawning grounds in the Gulf of Mexico (GOM) and Mediterranean Sea as well as differences in biological characteristics between fish from these regions (Fromentin and Powers, 2005; Rooker et al., 2007). Early genetic studies based on opportunistic sample collections were unable to detect population subdivision between putative stocks; however, subsequent genetic analyses that incorporated a biologically informed sampling design targeting larvae and mature adults on spawning grounds during the spawning season detected statistically significant population subdivision consistent with inferences from non-genetic data (Carlsson et al., 2007; Boustany et al., 2008).

Population genetic studies that incorporate sampling designs also inclusive of temporal replicates provide the ability to determine if an observation of statistically significant differentiation is stable over time and unlikely to result from artifacts such as the non-random sampling of populations, stochastic fluctuation in allele frequencies, or variation in reproductive success (Allendorf and Phelps, 1981; Waples and Teel, 1990; Hedgcock, 1994; Waples, 1998; Tessier and Bernatchez, 1999). Temporal replicates are particularly important for marine fishes because the level of genetic divergence among populations, if present, is expected to be low. In such cases, it becomes increasingly challenging to distinguish between noise and a weak but meaningful level of heterogeneity (Waples, 1998). Obtaining temporal replicates for some highly migratory marine fishes is especially challenging due to the relatively rare event nature of a number of these species. Frequently, these challenges result in very low sample sizes per sampling location in any given year, and necessitate the use of sample collections that are pooled across years for each location.

White marlin (Kajikia albida) is a highly migratory marine fish distributed throughout the Atlantic Ocean in temperate, subtropical, and tropical waters (Nakamura, 1985). This species is managed along with other pelagic fishes in the Atlantic Ocean by the member nations of the International Commission for the Conservation of Atlantic Tunas (ICCAT). Prior to the year 2000, ICCAT utilized a two stock management model for white marlin that recognized distinct populations in the North Atlantic and South Atlantic. This model was based on the presence of seasonally displaced spawning grounds north and south of the equator, fishery dependent data that reflected limited catches in equatorial waters, and a lack of trans-equatorial movements reported for tagged fish (ICCAT, 1994). ICCAT adopted a single stock management model in 2000 in response to new fisheries data that indicated continuous catches across the equator, trans-oceanic and trans-equatorial tag returns for a few individuals, and a population genetic study that was unable to detect statistically significant intra-oceanic heterogeneity (Graves and McDowell, 2001).

Since the adoption of the single stock management model for white marlin, additional genetic study has suggested the possibility of stock structure in this species. Analysis of variation at five nuclear microsatellite markers revealed statistically significant genetic differentiation between collections of white marlin from the western North Atlantic and western South Atlantic (WSA) ocean, and global levels of genetic differentiation that approached statistical significance based on the microsatellite data and on mitochondrial (mt) DNA control region sequence data (Graves and McDowell, 2006). The authors concluded that while the null hypothesis of genetic homogeneity could not be rejected, genetic population structure could exist but may not have been detected due to low statistical power and/or the opportunistic sampling design employed in the study (Graves and McDowell, 2006). Thus, there is considerable uncertainty regarding the suitability of a single stock management model for white marlin, and management of this species would benefit from a more thorough understanding of population structure.

In this study, the null hypothesis of a single Atlantic-wide stock of white marlin was evaluated by surveying genetic variation at 24 microsatellite loci and at the mtDNA control region in multiple collections of white marlin sampled from locations throughout the Atlantic Ocean. Relative to Graves and McDowell (2006), the ability to detect genetic population structure was improved by analyzing a larger number of microsatellite markers, increasing the number of sampling locations and sample sizes for some locations, and including a robust collection of larvae from the GOM.

Material and methods

Sample collection and generation of nuclear genotype data

Samples consisting of muscle tissue from landed adult white marlin or fin clips from live-released adult white marlin were opportunistically collected between 1992 and 2014 from the following locations: United States mid-Atlantic coast (USM, n = 263) off Cape May, NJ and Ocean City, MD; Caribbean Sea (CAR, n = 40) off the Dominican Republic and Cumaná, Venezuela; GOM (Adults, n = 49) off Veracruz, Mexico and from National Marine Fisheries Service pelagic longline survey stations throughout the GOM; western Central Atlantic (WCA, n = 55) off the northern and central coasts of Brazil; WSA (n = 39) off Santos, Brazil; and eastern North Atlantic (ENA, n = 33) off Morocco (Figure 1). Samples were stored at room temperature in 95% ethanol or a 10% DMSO solution. Biological information including sex, length, and weight was available for a limited number of samples; spawning condition was unknown for most specimens. White marlin larvae sampled from the northern GOM (Larvae, n = 75) between 2006 and 2013 were also evaluated; ichthyoplankton sampling methods are described in Rooker et al. (2012). Morphological species identification of larval white marlin was confirmed using the microsatellite genotype and mtDNA control region sequence data generated in this study, where larvae of species other than white marlin appeared as extreme outliers when compared with genotype and sequence data for adult white marlin.

Isolation of total genomic DNA was performed using standardized kits including the DNeasy Blood and Tissue Kit (Qiagen) and the ZR Genomic DNA Tissue MiniPrep Kit (Zymo Research). A subset of microsatellite loci previously developed for other istiophorid billfishes were optimized for use in white marlin (Buonaccorsi and Graves, 2000; Purcell et al., 2009; Sorensen et al., 2011; Bernard et al., 2012; Williams et al., 2015a). Twenty-four loci with consistent PCR amplification in white marlin (Supplementary Table S1) were developed into multiplex amplification reactions using forward primers modified at the 5' end
with locus-specific tag sequences and fluorescent dye labels. Amplifications were performed using the Type-It Microsatellite PCR Kit (Qiagen) following the manufacturer’s protocol. Amplification products were sized on an ABI 3130XL Genetic Analyzer (Applied Biosystems Inc.), and visualized using GeneMarker v2.6.0 software (SoftGenetics, LLC). Allele size calls were manually inspected for quality and a subset of samples (20%) were genotyped a second time to verify call consistency.

Generation of mtDNA sequence data
A segment of the mtDNA control region (~875 bp) was sequenced for a random subset of individuals from each sample collection and represented ≥ 29% of the total number of individuals in each collection (Table 1). PCR amplifications were performed using Pro-5 forward primer (Palumbi, 1996), an internal reverse primer (Graves and McDowell, 2006), and a PCR Core Reagents Kit (Qiagen). Amplification products were purified using the QIAquick PCR Purification Kit (Qiagen). Sequencing and consensus sequence construction were performed according to Portnoy et al. (2010), except with use of the MUSCLE multiple sequence alignment algorithm (Edgar, 2004). Control region sequence data previously generated for white marlin and reported in Graves and McDowell (2006) were downloaded from GenBank (accession numbers DQ835191–DQ835281) and included in the final alignment.

Analyses of microsatellite genotype data
Microsatellite genotype data were evaluated for the presence of scoring errors, null alleles, and large allele dropout using MicroChecker v2.2.3 (Van Oosterhout et al., 2004, 2006). Loci were tested for selective neutrality using the FST outlier detection method implemented in Lositan v1.0 (Antao et al., 2008) using 500 000 simulations, a stepwise mutation model, and neutral and reverse primer (Graves and McDowell, 2006), and a PCR Core Kit (Qiagen). Two scenarios of low and high genetic differentiation to produce the desired FST. Both simulation scenarios were completed with 1000 replications, effective population sizes of 1500, and 15 (low scenario) or 155 (high scenario) generations of drift. The proportion of significant results (defined as p < 0.05) was used to represent statistical power and was reported using Fisher’s exact test (Ryman and Palm, 2006). The probability of committing a Type I statistical error was also evaluated in each simulation.

Temporal replicates consisting of ≥ 10 individuals sampled from the same geographic location in more than 1 year were available for some sampling locations (Supplementary Table S3). These replicates were used to evaluate the consistency of a statistically significant population structure signal over time within a geographic region. Pairwise FST values were generated for temporal replicates within a sampling location using Arlequin. Significance of FST values was assessed based on 10 000 permutations of the data and a critical value corrected for multiple
pairwise comparisons (Benjamini and Yekutieli, 2001; Narum, 2006). The minimum number of samples required for a temporal replicate to be considered in this analysis (n = 10) was chosen based on the minimum number of samples associated with an acceptable (≥95% probability of detection) level of statistical power in power analyses performed using the high Fst scenario.

Analyses of mtDNA sequence data
Arlequin was used to determine the number of haplotypes and associated measures of haplotype and nucleotide diversity. Arlequin was also used to evaluate genetic differentiation globally across all samples, and pairwise between sample collections. The statistical significance of global and pairwise levels of genetic differentiation was assessed with 10,000 permutations of the data, and by correcting for multiple pairwise comparisons using a modified false discovery rate method (Benjamini and Yekutieli, 2001; Narum, 2006). A median joining haplotype network was generated in PopArt v1.7 (Leigh and Bryant, 2015).

Results
Summary statistics and genetic diversity
A total of 554 individuals representative of six geographic regions and two demographic groups were genotyped at 24 microsatellite loci (Supplementary Table S1). There was no indication of null alleles or large allele dropout in the genotype data. Genotype calls for the subset of samples genotyped twice were consistent between runs. One microsatellite locus (Isin29) demonstrated statistically significant deviation from the expectations of Hardy-Weinberg equilibrium in the WSA sample collection (p = 0.004). This deviation was associated with a deficiency of heterozygotes. As there was no pattern of deviation across multiple sample collections, Isin29 was retained for subsequent analyses. Observed and expected heterozygosity, including mean expected heterozygosity corrected for unequal sample sizes, were similar among sample collections (Supplementary Tables S1 and S2). No loci were identified as experiencing selection. The total number of alleles per locus ranged from 3 at Ta149 to 35 at Isin1 (mean a = 16.33); mean allelic richness within sample collections ranged from 9.96 to 10.39 and the mean number of private alleles per sample collection ranged from 0.23 to 0.46. Allelic richness for GOM Larvae (ar = 10.37; Supplementary Table S1) did not differ from allelic richness in any other sample collection (ar = 9.96 - 10.39), nor from the pooled collection of all adult samples (ar = 10.27).

An 858-bp alignment of the mtDNA control region was analysed for a total of 276 individuals, including 185 newly generated sequences (GenBank accession numbers K595230–K595414) and 91 sequences downloaded from GenBank (Table 1). Nucleotide composition consisted of 31.09% A, 29.37% T, 22.70% C, and 16.84% G. There were 354 polymorphic sites, 311 transitions, and 18 transversions. A total of 242 haplotypes were identified. Haplotype and nucleotide diversities across all samples were 0.99 and 0.031, respectively. Within sample collections, nucleotide diversity ranged from 0.014 to 0.036, and was lowest for USM (0.014) and CAR (0.016). Haplotype and nucleotide diversities in the larval collection (h = 1.00, π = 0.032; Table 1) were similar to that in all other sample collections (h = 0.99–1.00, π = 0.014–0.036) and to the pooled collection of all adult samples (h = 0.99, π = 0.031).

Population structure inferred from genetic data
Overall genetic differentiation based on the microsatellite genotype data were not statistically significant (global Fst = −0.00009, p = 0.525). Fst values associated with the pairwise comparison of sample collections ranged from 0 to 0.003 and were not statistically significant, with the exception of the comparison between the larval and adult sample collections from the GOM (Fst = 0.030, p = 0.010; Table 2). Results from STRUCTURE did not indicate the presence of more than one genetic group, and iterations with K = 1 were associated with the highest log likelihood values (Figure 2). STRUCTURE results were similar between admixture models that did or did not utilize sampling information as a prior to inform clustering. PCA also demonstrated a single grouping of individuals (Supplementary Figure S1). The eigenvalues associated with this analysis were similar across principal components.

Overall genetic differentiation based on the mtDNA sequence data were not statistically significant (global Fst = −0.00190, p = 0.614). Levels of genetic differentiation associated with pairwise comparisons of sample collections were not statistically significant (Table 3). The high haplotype diversity of the mtDNA control region was reflected in a midpoint spanning haplotype network (Figure 3). The three clusters of haplotypes apparent in this network were separated by 12 and 17 mutational differences.

Statistical power
Results from power simulations based on empirical allele frequency data generated in this study and in Graves and McDowell (2006) indicate that when the level of genetic differentiation between populations is relatively high (Fst = 0.05), an acceptable (≥95% probability of detection) level of statistical power is possible with low sample sizes (n ≥ 10) per population and either a small (n = 5) or large (n = 24) number of microsatellite markers

Table 1. Summary information for mtDNA control region sequence data.

<table>
<thead>
<tr>
<th>Sample collection</th>
<th>No. sequences</th>
<th>No. haplotypes</th>
<th>No. polymorphic sites</th>
<th>h</th>
<th>π</th>
</tr>
</thead>
<tbody>
<tr>
<td>WSA</td>
<td>17</td>
<td>17</td>
<td>134</td>
<td>1.00</td>
<td>0.036</td>
</tr>
<tr>
<td>WCA</td>
<td>40</td>
<td>39</td>
<td>166</td>
<td>0.99</td>
<td>0.031</td>
</tr>
<tr>
<td>USM</td>
<td>75</td>
<td>73</td>
<td>225</td>
<td>0.99</td>
<td>0.014</td>
</tr>
<tr>
<td>CAR</td>
<td>37</td>
<td>34</td>
<td>178</td>
<td>0.99</td>
<td>0.016</td>
</tr>
<tr>
<td>ENA</td>
<td>32</td>
<td>32</td>
<td>165</td>
<td>1.00</td>
<td>0.032</td>
</tr>
<tr>
<td>GOM Adults</td>
<td>44</td>
<td>41</td>
<td>185</td>
<td>0.99</td>
<td>0.032</td>
</tr>
<tr>
<td>GOM Larvae</td>
<td>31</td>
<td>31</td>
<td>176</td>
<td>1.00</td>
<td>0.032</td>
</tr>
<tr>
<td>Pooled Adults</td>
<td>245</td>
<td>216</td>
<td>347</td>
<td>0.99</td>
<td>0.031</td>
</tr>
<tr>
<td>All samples</td>
<td>276</td>
<td>242</td>
<td>354</td>
<td>0.99</td>
<td>0.031</td>
</tr>
</tbody>
</table>

h, haplotype diversity; π, nucleotide diversity.
Table 2. Matrix of pairwise $F_{ST}$ values based on microsatellite data (below diagonal) and pairwise $\phi_{ST}$ values based on mtDNA sequence data (above diagonal).

<table>
<thead>
<tr>
<th></th>
<th>WSA</th>
<th>WCA</th>
<th>USM</th>
<th>CAR</th>
<th>ENA</th>
<th>GOM Adults</th>
<th>GOM Larvae</th>
</tr>
</thead>
<tbody>
<tr>
<td>WSA</td>
<td>–</td>
<td>0.01796 (0.126)</td>
<td>0.01175 (0.167)</td>
<td>–</td>
<td>0.00206 (0.411)</td>
<td>–</td>
<td>0.00139 (0.398)</td>
</tr>
<tr>
<td>WCA</td>
<td>–0.00015 (0.447)</td>
<td>–</td>
<td>–0.00142 (0.471)</td>
<td>–</td>
<td>0.00561 (0.221)</td>
<td>–0.00386 (0.544)</td>
<td>–0.00445 (0.578)</td>
</tr>
<tr>
<td>USM</td>
<td>0.00006 (0.444)</td>
<td>–0.00125 (0.964)</td>
<td>–</td>
<td>0.01025 (0.109)</td>
<td>–0.00676 (0.769)</td>
<td>–</td>
<td>0.00285 (0.253)</td>
</tr>
<tr>
<td>CAR</td>
<td>–0.00118 (0.733)</td>
<td>–0.00066 (0.607)</td>
<td>–0.00053 (0.690)</td>
<td>–</td>
<td>–0.01313 (0.923)</td>
<td>–</td>
<td>–0.00283 (0.474)</td>
</tr>
<tr>
<td>ENA</td>
<td>0.00008 (0.440)</td>
<td>–0.00160 (0.783)</td>
<td>–0.00049 (0.643)</td>
<td>–0.00018 (0.448)</td>
<td>–</td>
<td>–0.00542 (0.596)</td>
<td>–0.01261 (0.904)</td>
</tr>
<tr>
<td>GOM Adults</td>
<td>0.00253 (0.059)</td>
<td>–0.00025 (0.478)</td>
<td>0.00128 (0.069)</td>
<td>0.00283 (0.041)</td>
<td>0.00134 (0.209)</td>
<td>–</td>
<td>–0.001496 (0.987)</td>
</tr>
<tr>
<td>GOM Larvae</td>
<td>–0.00092 (0.688)</td>
<td>–0.00166 (0.915)</td>
<td>–0.00035 (0.698)</td>
<td>–0.00054 (0.601)</td>
<td>0.00102 (0.222)</td>
<td>–</td>
<td>–0.00308 (0.010)</td>
</tr>
</tbody>
</table>

Significance values are shown in parentheses, statistically significant comparisons are italicized. A critical value corrected for multiple pairwise comparisons and equal to 0.014 was used.

Figure 2. STRUCTURE results based on microsatellite genotype data generated for 24 loci in white marlin in this study (A; plot shown for $K = 3$) and for 5 loci in striped marlin in McDowell and Graves (2008) (B; plot shown for $K = 4$). STRUCTURE results were similar with or without the use of sampling location as a prior to inform clustering. Individuals are organized by sample collection for both species. Sample collection labels for striped marlin are as follows: AUS, Port Stephens, Australia; CAL, San Diego, California; ECU, Manta, Ecuador; HAW, Kona, Hawaii; MEX, Cabo San Lucas, Mexico; TAW, Taiwan; and JAP, Japan.

Figure 3. Median joining haplotype network generated from mtDNA control region sequence data. Ancestral nodes inferred from observed data are shown in black. The number of mutational differences between nodes are represented by hatch marks along edges. Nodes are coloured according to sample collection, and node size corresponds to the number of occurrences for a particular haplotype.
equilibrium within all USM temporal replicates, with the exception of MnE in the 1995 replicate \((p = 0.008)\) and MnI in the 1998 replicate \((p = 0.011)\).

**Discussion**

The objective of this study was to evaluate genetic population structure in white marlin using a higher level of statistical power compared with previous studies. Increased statistical power was achieved by surveying a larger number of molecular markers across greater numbers of samples, including sample collections from additional geographic locations and a collection of larvae. With the increase in statistical power, levels of genetic heterogeneity in the present study decreased compared with those observed in previous studies with lower power. Graves and McDowell (2006) reported global levels of genetic differentiation that approached statistical significance \((F_{st} = 0.0022, p = 0.057; \phi_{st} = 0.0163, p = 0.069)\); however, in this study global levels of differentiation were considerably reduced \((F_{st} = -0.000099, p = 0.525; \phi_{st} = -0.00190, p = 0.614)\). The null hypothesis of a single genetic stock of white marlin could not be rejected based on the lack of genetic heterogeneity observed in this study.

This study represents the first population genetic analysis of an istiophorid billfish species to include a collection of larvae. Ideally, the inclusion of two or more collections of tissue from spawning adults or larvae sampled from geographically distinct spawning grounds would enable the direct evaluation of genetic differentiation between putative source populations of white marlin. However, confirmation of spawning status in adult fish requires direct observation of the release of gametes or the histological examination of gonadal tissue. Although the former approach may be possible for actively spawning females that may release hydrated oocytes when external pressure is applied to the ovaries, the latter approach requires the sacrifice of individual fish in order to inspect gonadal tissue. White marlin is an overfished species and the collection of fin clips as in this study prevents the need to sacrifice fish for the collection of tissues for genetic analysis. The collection of larvae from istiophorid billfishes is also challenging due to limited information on spawning in these species, and due to the logistics of obtaining robust sample sizes. In this study, sample collections of adult fish were not known to include actively spawning individuals, and only a single larval collection was available for genetic analysis.

Nevertheless, evaluation of a single collection of larvae can be informative of population structure when compared with collections of adults from throughout the species’ range. In the case of genetically discrete populations, a larval collection would be expected to display a statistically significant level of genetic differentiation when compared with collections of adults representative of another source population. In this study, there were no statistically significant levels of genetic differentiation between sample collections based on either the microsatellite or the mtDNA sequence data, with the exception of the microsatellite-based comparison of GOM Larvae and GOM Adults. The statistically significant heterogeneity observed between the GOM sample collections was driven by two loci (Ta149 and Isin29; Supplementary Figure S2) and resulted in an \(F_{st}\) for which the statistical significance was just below the critical value \((p = 0.010, p_{crit} = 0.014)\). This result was likely due to random sampling error and is not likely to be biologically meaningful (Waples, 1998). The lack of support for more than one genetic cluster in the STRUCTURE results is also consistent with the observed lack of genetic
results of this study lack any evidence to suggest the presence of more than one genetic stock of white marlin; however, a number of statistically significant pairwise comparisons were observed between temporal replicates from the WSA sample collection. These statistically significant comparisons are presumed to reflect random noise rather than a true population structuring signal. The WSA temporal replicates consisted of sample sizes ranging from 10 to 42 individuals per replicate; however, only replicates of very small sample sizes (1998, n = 11; 2012, n = 14) were associated with statistically significant pairwise comparisons. These results suggest that when statistical power is low due to small sample sizes and/or limited numbers of molecular markers, the ability to distinguish between noise and a true population structuring signal is diminished, resulting in a higher probability of detecting a false signal and committing a Type I statistical error (Waples, 1998). Conversely, low statistical power may also result in failure to detect a low but biologically meaningful population structuring signal and lead to a Type II statistical error.

The apparent lack of genetic population structure in white marlin has also been observed for other istiophorid billfishes. Previous evaluations of genetic population structure for blue marlin (Makaira nigricans) in the Atlantic Ocean have failed to detect population subdivision despite the analysis of multiple types of nuclear and mt markers (Buonaccorsi et al., 1999, 2001; McDowell et al., 2007). Genetic population subdivision was also not detected for sailfish (Istiophorus platypterus) in the Atlantic Ocean based on the analysis of mtDNA control region sequence data and a small number (n = 5) of microsatellite loci (McDowell, 2002; McDowell and Graves, 2002). Analysis of mtDNA control region sequence data for blue marlin and sailfish has demonstrated the presence of two distinct mt lineages for both species in the Atlantic Ocean (McDowell and Graves, 2002; McDowell et al., 2007). The existence of these lineages is attributed to historical isolation between Atlantic and Indo-Pacific populations of these species, followed by subsequent reintroduction of Indo-Pacific fish to the Atlantic Ocean (Graves and McDowell, 1995). In this study, haplotype diversity at the mtDNA control region is high, but distinct lineages are not present in white marlin. This result is consistent with a lack of historical isolation in white marlin compared with blue marlin and sailfish, perhaps due to differences in thermal tolerance and/or the spatial distribution of these species.

Despite the apparent lack of genetic population structure for some istiophorid billfish species in the Atlantic Ocean, genetic heterogeneity has been demonstrated for other highly migratory marine fishes in this ocean. Analyses of mtDNA control region sequence data and a number of nuclear loci, including microsatellite markers, revealed distinct populations of swordfish (Xiphius gladius) in the North Atlantic and South Atlantic (Chow and Takeyama, 2000; Alvarado-Bremer et al., 2005; Kasapidis et al., 2006). These populations correspond with the presence of distinct spawning grounds in the northern and southern hemispheres (Alvarado-Bremer et al., 2005). In the Pacific Ocean, genetic population structure has been reported for sailfish and black marlin (Istiophorus indica; McDowell, 2002; Lu et al., 2015; Williams et al., 2015b). In addition, the presence of at least four stocks has been genetically determined for striped marlin (Kajikia audax), the closely related sister species of white marlin, in the Pacific Ocean (McDowell and Graves, 2008; Purcell and Edmands, 2011). STRUCTURE analysis of genotype data previously generated for striped marlin (McDowell and Graves, 2008) and based
Genetic evaluation of population structure in white marlin

...on only five microsatellite loci clearly demonstrates the presence of four genetic clusters (Figure 2). Although Figure 2 provides an interesting comparison of the level of genetic subdivision observed for congeneric species that inhabit different ocean basins, the lack of populations resolved by STRUCTURE for white marlin may only reflect the limitations of this analysis in resolving subtle population structure in this species.

The genetic homogeneity observed for white marlin in this study may be consistent with what we currently know about the biology of this species. The distribution of catches based on fisheries-dependent data suggests a continuous species distribution in the Atlantic Ocean (ICCAT, 2015). Movements of white marlin inferred from the tagging of individual fish include a small number of conventional tag recaptures representing trans-oceanic (n = 7) or trans-equatorial (n = 3) dispersal (Snodgrass et al., 2013). Information from electronic tags reflects highly inconsistent directionality of movements by individuals tagged from seasonal assemblages of white marlin (Loose, 2014; Schlenker, 2014). In addition, seasonally displaced spawning grounds have been confirmed in the northern and southern hemispheres, including locations in the western North Atlantic, GOM, and Caribbean Sea for which spawning appears to primarily occur in the second quarter of the year (de Sylva and Breder, 1997; Luthy et al., 2005; Prince et al., 2005; Arocha and Barrios, 2009; Richardson et al., 2010; Rooker et al., 2012), and in the WSA where spawning primarily occurs in the fourth quarter (Ueyanagi et al., 1970; Arfelli et al., 1986; Amorim and Arfelli, 2003; Schmidt et al., 2015). Because the spawning season in these regions is not concurrent, it is possible that white marlin may spawn in multiple locations within a year, facilitating gene flow among geographically distant regions and resulting in the lack of genetic subdivision observed in this study.

The inability to reject the null hypothesis of a single genetic stock of white marlin in this study does not necessarily mean that ecological stocks of this species do not exist. Results of this study suggest that ocean-wide connectivity in white marlin is sufficient to prohibit the detection of genetic heterogeneity with the molecular markers surveyed in this study, but the magnitude of intraspecific connectivity beyond this threshold is unknown and could reflect dispersal in a small to large proportion of individuals. A 365-day geolocation track from Loose (2014) provides evidence that some white marlin complete cyclical migrations to seasonal foraging and spawning grounds on an annual time scale. Rooker et al. (2013, Supplementary Figure S5) also report cyclical movement for white marlin in the GOM based on a year-long geolocation track. Conventional tag recapture data also suggest annual periodicity associated with feeding and spawning assemblages of white marlin (Ortiz et al., 2003). If annual migrations are associated with some level of fidelity to discrete spawning grounds, this could result in the presence of ecological stocks; however, whether white marlin display spawning site fidelity is currently unknown. Atlantic-wide electronic tagging efforts that incorporate multi-year deployment periods would provide an informative approach for inferring ocean-wide connectivity in white marlin; however, tag deployment periods of this duration have not been previously successful.

Recently developed next-generation sequencing technology now facilitates the discovery of thousands of molecular markers representative of the whole genome, providing an unprecedented ability to detect genetic population structure (Davey and Blaxter, 2010; Narum et al., 2013). For example, application of this methodology to yellowfin tuna (Thunnus albacares; Grewe et al., 2015; Pecoraro et al., 2016) revealed inter- and intra-oceanic genetic heterogeneity previously unresolved by molecular markers such as allozymes, mtDNA, and microsatellites (Ward et al., 1994; Diaz-James and Uribe-Alcocer, 2006). Elucidation of unresolved or cryptic genetic heterogeneity can have significant consequences for fisheries management, particularly for stocks currently threatened by unsustainable fishing practices. The evaluation of genome-wide molecular markers may significantly improve the ability to detect genetic population structure, especially for weakly differentiated populations of marine species. In addition, genome-wide methodologies facilitate the identification of putative adaptive loci, which may provide insight on localized adaptation and regional genetic diversity, even if the level of intraspecific connectivity is enough to obscure heterogeneity at neutral markers (Allendorf et al., 2008; Andrews et al., 2016).

The ability to detect genetic population structure in white marlin was considerably improved in this study by increasing the number of molecular markers surveyed across a larger number of samples, including collections from additional geographic locations and larvae from the GOM. This increase in statistical power resulted in lower levels of genetic heterogeneity relative to those reported in a previous study that utilized fewer molecular markers and lower numbers of opportunistically collected samples. These results highlight the importance of statistical power in population genetic assessments of species for which population subdivision, if present, is expected to be shallow. Results from this study are consistent with the presence of a single genetic stock of white marlin in the Atlantic Ocean, and with the single-stock assessment and management model currently utilized by ICCAT. However, these inferences are based on molecular markers that may be limited in their ability to resolve low levels of genetic differentiation, and the degree of Atlantic-wide connectivity beyond that required to mask heterogeneity using the molecular markers surveyed in this study is unknown. It is especially important that Atlantic-wide connectivity in white marlin is understood considering the most recent stock assessment identified this species as overfished, with biomass in 2010 at half of that necessary to produce maximum sustainable yield (ICCAT, 2012). Additional investigation using genome-wide molecular markers and collections of larvae and/or reproductively active adults from additional spawning locations to inform inferences on ocean-wide genetic connectivity, as well as studies to determine the biological significance of the genetic results, are warranted.

Supplementary data

Supplementary materials are available at the ICESJMS online version of the article.

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