The Central Role of Dispersal in the Maintenance and Persistence of Seagrass Populations

Gary A. Kendrick
Michelle Waycott
Tim J. Carruthers
Marion :. Cambridge
Renae Hovey

See next page for additional authors

Follow this and additional works at: https://scholarworks.wm.edu/vimsarticles

Part of the Environmental Sciences Commons

Recommended Citation
Kendrick, Gary A.; Waycott, Michelle; Carruthers, Tim J.; Cambridge, Marion :.; Hovey, Renae; Krauss, Siegfried L.; Lavery, Paul S.; Les, Donald; Lowe, Ryan; Mascaro, Oriol; Ooi, Jillian S.; Orth, Robert J.; Rivers, David; Ruiz-Montoya, Leonardo; Sinclair, Elizabeth; Statton, John; Van Dijk, Jent Kornelis; and Verduin, Jennifer, "The Central Role of Dispersal in the Maintenance and Persistence of Seagrass Populations" (2012). VIMS Articles. 689.
https://scholarworks.wm.edu/vimsarticles/689

This Article is brought to you for free and open access by W&M ScholarWorks. It has been accepted for inclusion in VIMS Articles by an authorized administrator of W&M ScholarWorks. For more information, please contact scholarworks@wm.edu.
Authors
The Central Role of Dispersal in the Maintenance and Persistence of Seagrass Populations

GARY A. KENDRICK, MICHELLE WAYCOTT, TIM J. B. CARRUTHERS, MARION L. CAMBRIDGE, RENAE HOVEY, SIEGFRIED L. KRAUSS, PAUL S. LAVERY, DONALD H. LES, RYAN J. LOWE, ORIOL MASCARÓ I VIDAL, JILLIAN L. S. OOI, ROBERT J. ORTH, DAVID O. RIVERS, LEONARDO RUIZ-MONTOYA, ELIZABETH A. SINCLAIR, JOHN STATTON, JENT KORNELIS VAN DIJK, AND JENNIFER J. VERDUIN

Global seagrass losses parallel significant declines observed in corals and mangroves over the past 50 years. These combined declines have resulted in accelerated global losses to ecosystem services in coastal waters. Seagrass meadows can be extensive (hundreds of square kilometers) and long-lived (thousands of years), with the meadows persisting predominantly through vegetative (clonal) growth. They also invest a large amount of energy in sexual reproduction. In this article, we explore the role that sexual reproduction, pollen, and seed dispersal play in maintaining species distributions, genetic diversity, and connectivity among seagrass populations. We also address the relationship between long-distance dispersal, genetic connectivity, and the maintenance of genetic diversity that may enhance resilience to stresses associated with seagrass loss. Our reevaluation of seagrass dispersal and recruitment has altered our perception of the importance of long-distance dispersal and has revealed extensive dispersal at scales much larger than was previously thought possible.

Keywords: seagrass, long-distance dispersal, microsatellite DNA diversity, pollen, seed

The role of pollen and seed dispersal in the population dynamics and conservation of plant species is an emerging issue in plant ecology (Ouborg et al. 1999, Nathan 2006). Dispersal is fundamental to colonization, metapopulation dynamics, and plant migrations and will predictably be affected by human impacts and climate change in both terrestrial and aquatic environments (Cain et al. 2000). Interestingly, 41% of the world’s most widely distributed plant species are aquatic, despite those species’ representing fewer than 2% of all angiosperms (Cook 1996). Wide species distributions are attributed to efficient dispersal mechanisms and the capacity for long-distance dispersal (Les et al. 2003). Marine aquatic angiosperms are collectively known as seagrasses and are globally represented by a small number of species (72 species; Short et al. 2003). Seagrasses returned to the sea 100 million years ago, following the adaptation of their ancestors about 200 million years to terrestrial conditions (Les et al. 1997). These marine angiosperms are clonal plants that reproduce both vegetatively and sexually and that, with a combination of these reproductive modes, are able to persist and spread in dynamic nearshore environments. Important but unresolved areas in seagrass ecology include the influence of sexual reproduction in meadow maintenance, the way in which genetic variation can enhance resilience to stresses, and the role of dispersal in colonization of new or previously inhabited areas.

Historically, seagrass researchers have focused largely on vegetative reproduction, because it appeared to be responsible for extensive and long-lived meadows that allow species to withstand adverse conditions. As a result, seagrass conservation has been focused on the prevention of losses of vegetative material or on the adoption of restoration strategies that use vegetative cuttings from existing meadows to ameliorate site disturbances (Paling et al. 2009). The role of other life-history stages has largely been ignored, with some exceptions (Orth et al. 1994, 2000, 2006a); however, the large investment of energy in producing flowers, fruits, and seeds and the high levels of genetic diversity among seagrass populations suggest that sexual reproduction contributes to the persistence and maintenance of meadows. Research on restoration techniques that use seeds to promote the successful recolonization of habitats is also justified (e.g., Marion and Orth 2010) and is critical to creating resilient, genetically diverse seagrass habitats.

Studies on other aquatic plants indicate that seed dispersal, in particular, plays a key role in shaping the distribution, structure, and resilience of aquatic plant populations (Les et al. 2003). Emerging results of molecular studies of...
seagrasses support the claim that seed dispersal has a role in contributing to the genetic complexity of seagrass habitats, with high overall genotypic diversity and weak genetic structuring at large spatial scales within species. In the present article, we review the dispersal mechanisms of seagrasses, with a focus on their reproductive biology, adaptation to water pollination (hydrophily), and seed dispersal. We explore the patterns in spatial genetic variation that may give significant insight into long-distance dispersal and consider the implications that these patterns have on species distribution, habitat diversity, and resilience to environmental stress. As a result of our evaluation, we contend that sexual reproduction and dispersal are critically important to the ecological conservation and restoration of seagrasses. An awareness of these implications is essential for informing appropriate conservation and management strategies.

Introducing seagrasses

Seagrasses are submerged aquatic flowering plants that are not directly related to terrestrial grasses. Seagrasses have evolved within the monocotyledon angiosperm group known as the Alismatidae (Les et al. 1997, Waycott et al. 2006). There are three or four separate major evolutionary events that resulted in contemporary seagrasses, which are currently placed within six taxonomic families (Waycott et al. 2006). The 72 seagrass species represent around two-hundredths of one percent of all flowering plants (Short www.biosciencemag.org 2008). Such unisexual flowers present further challenges for successful pollination and seed set among the angiosperms as a whole (4% of species; Richards 1986). Such unisexual flowers present further challenges for successful pollination and seed set, because pollen must reach small and often distant receptive female stigmas via a highly inefficient vector (water currents).

Once established, seagrass meadows are able to expand through vegetative clonal growth by rhizome extension. As a result, individual plants that arise from seed (genets) have the capacity to form large clones comprising many often physiologically disconnected shoots (ramets). In this respect, seagrasses are similar to many other rhizomatous terrestrial and aquatic monocotyledons. Because seagrasses have two major reproductive strategies (i.e., both sexual and vegetative), the formation of established seagrass meadows typically requires that the two strategies act in concert. For example, seeds or seedlings would logically represent the primary mode of dispersal into new, uncolonized locations. However, once recruitment has occurred, the plants typically expand by rhizomatous growth. Therefore, the production and dispersal of sexual propagules and the subsequent establishment of seedlings are critical life-history stages for seagrasses. Despite their importance, these processes have received relatively little attention in studies in which the global resilience and survival of seagrass habitats were addressed (Waycott et al. 2006).

Reproduction and flowering

Seagrass flowers develop within or above the canopy or at or below the sediment surface (figure 1). Flowers in the canopy are generally more robust than those at the sediment surface, which probably indicates an adaptation to hydrodynamic (wave and current) forces. Like many perennial plants, seagrasses must achieve a balance between resources allocated to flowering and those allocated to the ongoing production and maintenance of vegetative tissues (Obeso 2002). Although relative resource allocation to sexual reproduction has been poorly studied in seagrasses, the large variability in flowering rates and seed set among seagrass populations implies that some type of trade-off does occur (figure 2).

Flowering, much like vegetative growth, is regulated by temperature, light, and nutrients. For example, the temperate Australian species of Posidonia all typically flower in midwinter (Smith and Walker 2002), when water temperatures are lowest, day lengths are shortest, and storm frequency is highest. However, abnormally high sea temperatures can alter this pattern dramatically by inducing the onset of flowering in Posidonia oceanica (Diaz-Almela et al. 2006). Herbivory also appears to influence the timing of anthesis in some seagrass species. In the subtidal Thalassia testudinum, pollen is released within an hour of dusk, possibly to escape grazing by parrotfish (van Tussenbroek et al. 2008). The relationship between the onset of reproduction and pollen dispersal and storm frequency and the potential link between hydrodynamics and increased pollen-dispersal distances has not yet been studied quantitatively.

Pollen dispersal in seagrasses

Pollen dispersal into seawater could provide a mechanism of long-distance dispersal, somewhat similar to that observed for wind-dispersed pollen. Unlike wind-dispersed pollen in terrestrial plants, seagrass pollen is generally very sticky and readily adheres to any surface that it encounters. In either case, pollen grains need to occur in sufficient numbers and proximity to the stigmas of the female flowers in order to enable successful reproduction (figure 1). The efficacy of seagrass pollination depends on the relative distribution of males and females within the population (i.e., the relative spatial distribution of genets); the morphology, viability, and adhesiveness of the pollen; stigmatic modifications that facilitate pollen capture; the location of pollen release (with respect to female flowers) in the canopy; canopy structure; and various hydrodynamic interactions (Inglis and Waycott...
that release their pollen only after encountering a female flower also at the water surface (Les et al. 1997, Tanaka et al. 2004).

The extent of pollen-dispersal distances is determined in part by the position of the flowers in the seagrass canopy (figure 1a; Verduin et al. 2002). Pollen released at the base or at midcanopy is entrained locally within the meadow; therefore, there is a high probability of local mating success and a lower probability of long-distance dispersal. Several seagrasses (Posidonia australis, Syringodium, Phyllospadix, and Zostera) produce flowers toward the top of or even above the canopy (Ackerman 2006), where pollen and occasionally whole anthers are released into the water column (e.g., Amphibolis; McConchie and Knox 1989). The release of pollen above the canopy should facilitate the long-distance dispersal of some pollen, although much of the pollen is likely to become entrained locally because of synchronous leaf fluttering (known as monami) in a seagrass canopy in response to waves and currents (Grizzle et al. 1996).

The potential of seagrass pollen for long-distance dispersal is enabled by its viability, which can extend to over 50 hours for Posidonia (Smith and Walker 2002) and to at least 8 hours for Zostera (Cox et al. 1992). At the upper end of this range, the period of pollen viability is comparable to that in other plants with long-distance dispersal: Insect-pollinated terrestrial plants can retain pollen viability for 36–48 hours, and some wind-pollinated terrestrial plants retain viability for more than 72 hours (e.g., Pacini et al. 1997).

However, seagrass pollen is not ideally designed for long-distance dispersal. The large size (up to 0.5 millimeters; Knox 1984) and elongate shapes that often characterize the pollen of subsurface pollinated species are not conducive to
distant transport in water, and the rafting chains of floating pollen rapidly disperse by wave action as the distance from their release point increases (Ackerman 1995). On the basis of its architecture, seagrass pollen appears better suited for local dispersal within meadows. The higher level of dioecy observed in seagrasses presumably functions to reduce within-genet pollination that might otherwise predominate as a result of near-neighbor pollination. Studies in mating systems and paternity assignment in Zostera noltii and Zostera marina suggest that, despite very high outcrossing rates, these species’ pollen travels a maximum distance of 15 meters (m), which suggests that local environmental characteristics play a major role in dispersal (Zipperle et al. 2011).

**Dispersal potential of seeds and propagules**

The dispersal of seagrass propagules occurs through abiotic transport by ocean waves and currents, through sediment movement, or potentially through transport by biotic vectors like manatee, dugong, turtles, and geese (figure 1). Similarly, the wide distributions of a large number of nonmarine aquatic plants have been achieved primarily through the transport of propagules by highly vagile birds, which are known to disperse seagrasses (Les et al. 2003). Terrestrial plants have a much wider array of biotic and abiotic dispersal agents available.

The response of seagrass propagules to waves and currents at a location will depend largely on the physical properties (especially buoyancy) of the seed, fruit, or vegetative bract (spatha) that surrounds the seeds (figure 2). Seagrass propagules are produced as two basic forms: positively buoyant (floating) fruit, spathe, and viviparous seedlings and neutrally or negatively buoyant seeds and viviparous seedlings. The seeds of seagrasses are generally neutrally or negatively buoyant, because they must settle on the substrate at considerable depth. However, seeds can be retained within positively buoyant structures such as fruits or spathe or can germinate as viviparous seedlings. All of these structures eventually sink, sometimes after being transported considerable distances.

Species that do not form floating structures release their seeds within the mother meadow and are dispersed locally by turbulence or rely on ingestion by herbivores (e.g., manatee, dugong, turtle) for dispersal over greater distances (figure 1). These different types of dispersal are well represented among seagrass genera (figure 2). Posidonia, Enhalus, and Thalassia produce floating fruits; Zostera and Syringodium produce floating rhiopia, spathe, and seeds; and Thalassodendron produces floating viviparous seedlings. Cymodocea, Halophila, and Halodule produce seeds that are neutrally or negatively buoyant, and Amphibolis produces negatively buoyant viviparous seedlings.

The farthest measured dispersal distances (300–400 kilometers [km]) have been recorded for those seagrass genera with floating fruit and rhiopia (figure 3; e.g., Enhalus and Thalassia). Floating may be viewed as a long-distance dispersal strategy in cases in which dispersal distances are influenced by regional ocean-current systems. For example, the floating fruits of *T. testudinum* can be transported for more than 300 km when they are captured by offshore currents (van Dijk et al. 2009).

For negatively buoyant propagules, the potential scale of dispersal is limited by hydrodynamic processes occurring within the bottom boundary layer (figure 1b)—that is, those processes required for the vertical mobilization of propagules into the water column, where they can be transported horizontally by unidirectional currents. Interestingly, once they have settled onto the sediment surface, the seeds of *Zostera* require significant energy to be transported farther (Harwell and Orth 2002). However, even though primary seed dispersal may be localized when the seeds are in the sediment, there is the potential for secondary dispersal as sediments are resuspended and transported under high-energy storm conditions. Secondary dispersal may also occur through transport by herbivores like turtles, dugong, manatee, ducks and geese, although the viability of the seeds after ingestion requires further study (figure 1c, 1d). In *Halophila*, the seeds have been shown to be moved by bulk sediment transport during tropical storms and may also be transported in the guts of grazing dugongs and turtles (e.g., Heck et al. 2008).

The mobilization of negatively buoyant propagules also occurs when the shear stress at the bed (or at the canopy) exceeds some critical threshold. In coastal systems, where some wave energy is typically present, shear stresses induced by oscillatory wave motion are generally much greater (by a factor of 10–100) than those induced by unidirectional currents with equivalent flow speeds. Therefore, the dispersal distances of these propagules may ultimately depend on the amount of local wave energy available at a given site, and major dispersal events probably occur only during extreme wave events—for example, during cyclones (Hammerstrom et al. 2006). Species like *Cymodocea nodosa*, with seeds that are negatively buoyant or buried in the sediment, can persist in highly dynamic environments through recolonization of the substrate by the seeds after a perturbation (Marbà and Duarte 1995).

Conversely, the dispersal distance of positively buoyant propagules is most strongly influenced by processes occurring within the surface boundary layer (figures 1b and 2)—that is, subsequent to their release up to the surface of the water column. The floating fruits of *Thalassia* and *Enhalus* (e.g., Lacap et al. 2002, van Dijk et al. 2009), as well as the floating reproductive shoots of *Zostera* (e.g., Erftemeijer et al. 2008, Källström et al. 2008), can attain large dispersal distances as a result of near-surface transport processes, especially when these processes are driven by local wind stresses. Although these floating propagules are transported directly by surface water currents, their transport rates also are influenced by winds (windage effects), which exert drag forces (Erftemeijer et al. 2008, van Dijk et al. 2009).

The direct transport of floating fruit or rhiopia by surface wave motions (e.g., Stokes drift) has yet to receive attention in the
seagrass literature; however, it could represent an important (or even dominant) transport mechanism in wave-dominated coastal environments.

Overall, long-distance dispersal in seagrasses appears far more likely to occur through seeds, fruits, viviparous seedlings, and vegetative fragments than by pollen. These structures have the capacity to move distances ranging from hundreds of meters to hundreds of kilometers. Seagrass dispersal has been directly and empirically addressed in few studies. In studies on *Z. marina*, Harwell and Orth (2002), Lacap and colleagues (2002), and Källström and colleagues (2008) concluded that long-distance dispersal to distances of 20–300 km were possible. In *T. testudinum*, van Dijk and colleagues (2009) similarly found that although the majority of the fruits in a meadow released their seed close to the parent plant, floating fruits occasionally had a much higher dispersal capacity, with distances of up to 8 km (Kinlan and Gaines 2003). Long-distance dispersal of vegetative fragments is possible, and the natural recruitment by dispersed floating vegetative fragments (which include rhizome fragments) has been reported (Di Carlo et al. 2005, Hall et al. 2006). However, few researchers have evaluated how effectively such floating, vegetative fragments can reestablish.

**Measuring dispersal in seagrasses with molecular markers**

Molecular markers have provided several insights into the population genetic structure of seagrasses from which patterns and mechanisms of dispersal can be inferred. Population genetic studies have shown a trend whereby meadows at the extremes of a species range tend to show less genetic diversity (Waycott et al. 1997, Olsen et al. 2004, Arnaud-Haond et al. 2007, van Dijk et al. 2009). High genotypic diversity exists at all spatial scales within most species (table 1, and see the supplemental appendix S1, available online at [http://dx.doi.org/10.1525/bio.2012.62.1.10](http://dx.doi.org/10.1525/bio.2012.62.1.10)). The full range of within-meadow genotypic diversity (from a single clone to no clonality detected) exists among seagrass meadows, even within close proximity (Ruggiero et al. 2002, Muñiz-Salazar et al. 2005, 2006, Alberto et al. 2006). Consequently, the processes affecting the dispersal of pollen, seed, and vegetative fragments at smaller spatial scales are complex and variable. The fixation index ($F_{ST}$) and theta ($\theta_{ST}$) measure genetic subdivision among populations, or genetic diversity. Low levels of spatial genetic subdivision (i.e., measured using the fixation index $F_{ST} < 0.1$) are typical at local scales (less than 100 km; see table 1). These observations of high genetic diversity in many seagrass meadows and weak
spatial genetic structuring among meadows suggest that sexual reproduction in seagrasses is more important than was previously thought and that seagrass population structure is not always dominated by clonal processes.

Patterns of spatial genetic variation in widespread seagrasses assessed over broad geographical ranges have also revealed significant insights into long-distance dispersal. Extensive, range-wide genetic studies typically show pairwise population genetic distances increasing proportionally with geographical distance at broad scales (hundreds of kilometers) but show weak relationships or no significant relationship at more regional scales (less than 200 km) (figure 4a–4d). Significantly, these genetic data strongly suggest a general threshold long-distance dispersal distance at hundreds of kilometers for widespread seagrass species—a distance that coincides with some directly observed seed-dispersal distances (Harwell and Orth 2002).

Table 1. Genetic diversity indices and sampling scales for seagrass species using microsatellite DNA markers.

<table>
<thead>
<tr>
<th>Species</th>
<th>Number of meadows</th>
<th>Sampling scale</th>
<th>Geographic sampling range (km)</th>
<th>Number of ramets</th>
<th>Number of genets</th>
<th>Number of genets/ramets</th>
<th>Mean $H_o$</th>
<th>Mean $H_e$</th>
<th>AMOVA or global $F_{ST}$</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cymodocea nodosa</td>
<td>10</td>
<td>Regional</td>
<td>15–300</td>
<td>373</td>
<td>255</td>
<td>0.68</td>
<td>—</td>
<td>0.43</td>
<td>0.18</td>
<td>Alberto et al. 2006</td>
</tr>
<tr>
<td>Cymodocea nodosa</td>
<td>47</td>
<td>Entire range</td>
<td>5500</td>
<td>1784</td>
<td>789</td>
<td>0.44</td>
<td>0.48</td>
<td>0.48</td>
<td>0.43</td>
<td>Alberto et al. 2008</td>
</tr>
<tr>
<td>Thalassia testudinum</td>
<td>16</td>
<td>Regional</td>
<td>1350</td>
<td>573</td>
<td>315</td>
<td>0.68</td>
<td>0.66</td>
<td>0.10</td>
<td></td>
<td>van Dijk and van Tussenbroek 2010, van Dijk et al. 2009</td>
</tr>
<tr>
<td>Posidonia australis</td>
<td>12</td>
<td>Local</td>
<td>80</td>
<td>587</td>
<td>348</td>
<td>0.59</td>
<td>0.49</td>
<td>0.48</td>
<td>0.11</td>
<td>Serra et al. 2010</td>
</tr>
<tr>
<td>Posidonia oceanica</td>
<td>27</td>
<td>Entire range</td>
<td>3500</td>
<td>919</td>
<td>650</td>
<td>0.71</td>
<td>0.45</td>
<td>—</td>
<td>0.24</td>
<td>Arnaud-Haond et al. 2007</td>
</tr>
<tr>
<td>Posidonia oceanica</td>
<td>34</td>
<td>Entire range</td>
<td>3500</td>
<td>1217</td>
<td>875</td>
<td>0.72</td>
<td>0.43</td>
<td>—</td>
<td>0.23</td>
<td>Diaz-Almela et al. 2006</td>
</tr>
<tr>
<td>Posidonia oceanica</td>
<td>4</td>
<td>Entire range</td>
<td>3500</td>
<td>307</td>
<td>218</td>
<td>0.71</td>
<td>—</td>
<td>—</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Zostera marina</td>
<td>6</td>
<td>Local</td>
<td>53</td>
<td>485</td>
<td>485</td>
<td>1.00</td>
<td>0.50</td>
<td>0.53</td>
<td>0.02</td>
<td>Oetjen and Reusch 2007</td>
</tr>
<tr>
<td>Zostera marina</td>
<td>6</td>
<td>Local</td>
<td>53</td>
<td>284</td>
<td>284</td>
<td>1.00</td>
<td>—</td>
<td>0.36</td>
<td>0.02</td>
<td>Oetjen et al. 2010</td>
</tr>
<tr>
<td>Zostera marina</td>
<td>12</td>
<td>Local</td>
<td>~80</td>
<td>281</td>
<td>242</td>
<td>0.86</td>
<td>0.61</td>
<td>0.62</td>
<td>0.14</td>
<td>Tanaka et al. 2011</td>
</tr>
<tr>
<td>Zostera marina</td>
<td>13</td>
<td>Regional</td>
<td>442</td>
<td>390</td>
<td>301</td>
<td>0.77</td>
<td>0.47</td>
<td>0.47</td>
<td>0.17</td>
<td>Becheler et al. 2010</td>
</tr>
<tr>
<td>Zostera marina</td>
<td>9</td>
<td>Regional</td>
<td>220–2300</td>
<td>328</td>
<td>328</td>
<td>1.00</td>
<td>0.55</td>
<td>0.57</td>
<td>0.16</td>
<td>Muñiz-Salazar et al. 2005</td>
</tr>
<tr>
<td>Zostera marina</td>
<td>12</td>
<td>Entire range</td>
<td>12–10000</td>
<td>545</td>
<td>371</td>
<td>0.68</td>
<td>0.47</td>
<td>0.48</td>
<td>0.29, 0.49</td>
<td>Muñiz-Salazar et al. 2005</td>
</tr>
<tr>
<td>Zostera marina</td>
<td>49</td>
<td>Entire range</td>
<td>10–10000</td>
<td>2139</td>
<td>1438</td>
<td>0.67</td>
<td>0.43</td>
<td>0.44</td>
<td>0.40</td>
<td>Oetjen et al. 2010</td>
</tr>
<tr>
<td>Zostera marina</td>
<td>8</td>
<td>Regional</td>
<td>60–10000</td>
<td>372</td>
<td>233</td>
<td>0.63</td>
<td>0.56</td>
<td>0.57</td>
<td>0.18</td>
<td>Diekmann et al. 2005</td>
</tr>
<tr>
<td>Zostera noltii</td>
<td>33</td>
<td>Entire range</td>
<td>2–8000</td>
<td>1706</td>
<td>1118</td>
<td>0.66</td>
<td>0.53</td>
<td>0.52</td>
<td>—</td>
<td>Coyer et al. 2004</td>
</tr>
<tr>
<td>Zostera pacifica</td>
<td>17</td>
<td>Regional</td>
<td>~300</td>
<td>783</td>
<td>291</td>
<td>0.37</td>
<td>—</td>
<td>0.13</td>
<td>—</td>
<td>Coyer et al. 2008</td>
</tr>
</tbody>
</table>

Note: Entire range represents the entire geographic distribution of the species. AMOVA, analysis of molecular variance; $H_o$, observed heterozygosity; $H_e$, expected heterozygosity; $F_{ST}$, measure of genetic subdivision; km, kilometers.
van Dijk et al. 2009). Similarly, genetic data for *Z. marina* (Olsen et al. 2004) supported predictions based on experimental modeling (Källström et al. 2008), which indicated that dispersal distances of as much as 150 km are possible for this species.

The spatial distribution of genetic variation within and among populations of seagrasses provides considerable insight into dispersal vectors at smaller spatial scales (e.g., Waycott et al. 2006, van Dijk and van Tussenbroek 2010). For example, in *T. testudinum*, the occurrence of identical shared multilocus genotypes over 230 m suggests that rhizomes can extend over large distances or that vegetative fragments can successfully recruit (van Dijk and van Tussenbroek 2010). In contrast, for *Posidonia australis*, no multilocus genotypes (clones) were shared among sampled meadows along an 80-km stretch of coastline (*N* = 587), which supports the suggestion that sexually produced seeds, rather than vegetative fragments, are the main source of long-distance dispersal.

In terrestrial plants, the vast majority of seeds disperse close to their parent plant, and the very few long-distance dispersal events are usually due to “non-standard” vectors that are different from those responsible for local dispersal (Nathan 2006). Although it remains unclear whether a similar pattern applies to marine environments, where habitats can be more highly connected, a fundamental difference from terrestrial plant dispersal is that the standard vector of ocean currents remains responsible for both local dispersal and long-distance dispersal. In addition, it might be expected that stochasticity associated with long-distance dispersal, which is high in terrestrial plants (Nathan 2006), is substantially lower in
seagrasses, leading to more regular dispersal events and an erosion of spatial genetic structure at large spatial scales (figure 4). The maximum dispersal distances in sedentary marine species appear to exceed the maximum estimates for terrestrial plants by at least one to two orders of magnitude (Kinlan and Gaines 2003). Although seagrasses were not considered by Nathan (2006) or by Kinlan and Gaines (2003), patterns of broad-scale genetic structure for widespread seagrass species (figure 4) support this observation.

The recent development of gene-linked microsatellites derived from expressed sequence tag libraries (EST-microsatellites) in Z. marina and P. oceanica (Wissler et al. 2009) highlight exciting future opportunities to further assess the genetic importance of long-distance dispersal and, therefore, the ability of species to shift ranges, as well as their genetic capacity to adapt to changing environmental conditions. For example, genome scans in Z. marina have shown divergent selection among subtidal versus intertidal meadows, showing evidence for habitat-specific selection (Oetjen and Reusch 2007, Oetjen et al. 2010), with consequences for realized dispersal in the face of rapid climate change.

**Implications for managing disturbance to seagrass ecosystems**

Seagrasses inhabit some of the most threatened coastal ecosystems in the world (Orth et al. 2006b, Waycott et al. 2009), and a more thorough understanding of the nature and extent of their dispersal is essential for implementing appropriate management and restoration strategies under predominantly anthropogenic conditions (Orth et al. 2006b). A better understanding of the relationships between the frequency and the range of pollen, seed, and propagule dispersal, with respect to particular spatial scales of disturbance, should help managers decide whether to actively enhance recruitment into a disturbed area (e.g., rehabilitation) or to manage an area for natural recruitment. In genera such as Halophila, existing seed banks can facilitate recovery following a disturbance (Marbà and Duarte 1995) and should allow natural recovery from small- to medium-scale disturbances, as long as the habitat quality (such as water quality, sediment quality, and geomorphology) is maintained. Conversely, genera such as Posidonia, which flower only once a year and do not form seed banks, are characterized by a small window of recruitment opportunity and would require more active management, through restoration, to remediate losses. In cases in which restoration would involve the deliberate introduction of propagules, it is important to understand patterns of genetic diversity and dispersal thoroughly in order to determine the most appropriate provenance for propagule sources. In cases in which seed dispersal is widespread and genetic subdivision is low among populations, seed stocks representing a broad provenance should have minimal negative restoration consequences. The development of restoration methods using seeds provides a better opportunity to maintain genetic diversity within restored sites while dramatically reducing the impact on donor sites.

**Conclusions**

Seagrasses represent several specialized groups of monocotyledons, which have adapted successfully to colonize the shallow coastal seas and estuaries of the world. This review has described several unique aspects of flowering and pollination, the dispersal of reproductive propagules, and seedling recruitment across this group. The flowers occur at the sediment surface, within the canopy, and above the canopy. Seagrass pollen is among the largest for all angiosperms and is modified for water pollination in its shape, buoyancy characteristics, and size. In many seagrasses, pollination appears to occur predominantly at a local scale, within meadows, because most pollen is trapped within the canopy when released. In species that release pollen above the canopy, the potential for long-distance dispersal is enhanced. Ultimately, the power of molecular markers for paternity studies is required for a true understanding of the realized patterns of pollen dispersal within seagrasses.

Seagrasses possess diverse dispersal mechanisms that include floating fruits, rhipidia, and viviparous seedlings that are capable of long-distance dispersal and can potentially be transported over considerable distances in marine systems. In contrast, some seagrass species produce their seeds within the sediment or on the sediment surface, and dispersal away from the parental plants is minimal unless the sediments as a whole are moved by major disturbances (e.g., cyclones and extreme storms). Similar processes have been described for terrestrial plants (Nathan et al. 2008).

Our reevaluation of seagrass dispersal and recruitment has altered our fundamental understanding of seagrass resilience in several ways. It is evident that seagrasses are resilient and have persisted in a physiologically challenging submerged environment because they have broad realized niches. However, it now is apparent that the local persistence of seagrasses has been achieved not only by clonal growth but also by recruitment from sexually derived propagules. Seagrasses invest significant amounts of energy in sexual reproduction producing seeds with a high capacity for long-distance dispersal that enables them to colonize distant new locations. Although the frequency, extent, influences, and consequences of long-distance dispersal are still poorly known for many seagrass species, recent assessments of range-wide population genetic variation in widespread seagrass species have revealed extensive dispersal at scales much larger than was previously thought possible. Molecular studies in combination with ecological and hydrological assessments offer exciting new opportunities for a more-detailed understanding of long-distance dispersal in seagrasses.

**Acknowledgments**

This article was based on discussions from the ARC-NZ Research Network for Vegetation Function Working Group.
References cited


Gary A. Kendrick (gary.kendrick@uwa.edu.au) is affiliated with the Oceans Institute and with the School of Plant Biology, both at the University of Western Australia, in Crawley. Michelle Waycott is affiliated with the School of Earth and Environment, The University of Adelaide, South Australia, Australia. Tim J. B. Carruthers is affiliated with the Integration and Application Network at the University of Maryland Center for Environmental Science, in Cambridge, and with the Secretariat of the Pacific Regional Environment Programme in Apia, Samoa. Marion L. Cambridge is affiliated with the Oceans Institute and with the School of Plant Biology, both at the University of Western Australia, in Crawley. Renee Hovey is affiliated with the Oceans Institute and with the School of Plant Biology and Earth and Environment, at the University of Western Australia, in Crawley. Siegfried L. Krauss is affiliated with the School of Plant Biology at the University of Western Australia, in Crawley, and with Kings Park and Botanic Garden, under the Botanic Gardens and Parks Authority, in West Perth, Western Australia. Paul S. Lavery is affiliated with the Centre for Marine Ecosystems Research and School of Natural Sciences at Edith Cowan University, in Joondalup, Western Australia. Donald H. Les is affiliated with the Department of Ecology and Evolutionary Biology at the University of Connecticut, in Storrs. Ryan J. Lowe is affiliated with the Oceans Institute and with the School of Earth and Environment, both at the University of Western Australia, in Crawley. Orval Mascaró i Vidal is affiliated with the Department of Ecology at the University of Barcelona, in Spain. Jillian L. S. Ooi is affiliated with the Oceans Institute, the School of Plant Biology, and the School of Earth and Environment at the University of Western Australia, in Crawley, and with the Department of Geography at the University of Malaya, in Kuala Lumpur. Robert J. Orth is affiliated with the School of Marine Science and the Virginia Institute of Marine Science at the University of Virginia, in Gloucester Point, Virginia. David O. Rivers is affiliated with the Oceans Institute and School of Plant Biology at the University of Western Australia, in Crawley. Leonardo Ruiz-Montoya is affiliated with the Oceans Institute, the School of Plant Biology, and the School of Earth and Environment at the University of Western Australia, in Crawley. Elizabeth A. Sinclair is affiliated with the School of Plant Biology at the University of Western Australia, in Crawley, and with Kings Park and Botanic Garden, under the Botanic Gardens and Parks Authority, in West Perth, Western Australia. John Stanton is affiliated with the Oceans Institute and with the School of Plant Biology, both at the University of Western Australia, in Crawley. Jennifer Verduin is affiliated with the School of Environmental Science at Murdoch University, in Perth, Western Australia.