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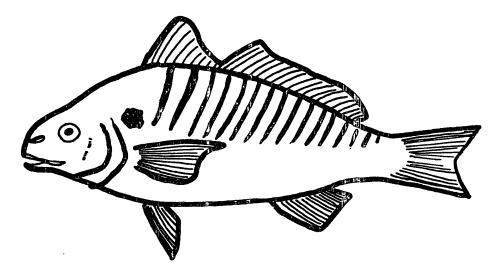
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A LABORATORY ANALYSIS OF KEPONE DEPURATION BY SPOT, LEIOSTOMUS XANTHURUS

by Marion Y. Hedgepeth, Robert T. Doyle, and Linda L. Stehlik



Special Report In Applied Marine Science and Ocean Engineering #229

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ACKNOWLEDGEMENTS

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INTRODUCTION

Dectectable residue levels of the pesticide Kepone have been found in resident and migratory finfishes from the James River, Virginia (Bender et al., 1977). As a result, the James River was closed to commercial finfishing in early 1976 (with the exceptions of channel catfish and American shad for a short period of time). In addition, the United States Food and Drug Administration established an action level of 0.3 ppm of Kepone in finfishes utilized for human consumption.

Residue levels of Kepone in fishes such as spot, Atlantic croaker, bluefish, striped bass and American shad were investigated to determine if these migratory species present a health hazard to the public in areas beyond the James River system. Bender et al. (1977) found that residue levels in finfishes were dependent upon the species of fish and the length of residence in the James River. Also, they maintained that residue levels in finfishes declined as distance from the James was increased.

In 1977, additional Kepone studies were begun at the Virginia Institute of Marine Science (VIMS) to determine the rates of Kepone depuration in contaminated fishes from the James River. In a laboratory analysis of Kepone depuration by Atlantic croaker, <u>Micropogonias undulatus</u>, Doyle et al. (In Press) observed a significant drop in Kepone concentration in the 24th week sample. Furthermore, it was noted that this significant change in mean residue levels coincided with a rise in the ambient water temperature to above 15°C; however, additional studies were needed to confirm this relationship. In our study we chose to observe the effect of temperature on the rate of Kepone depuration by contaminated spot, Leiostomus xanthurus, from the James River.

Bender et al. (1977) reported a mean Kepone level of 0.81 ppm in spot from the James River and the lower Chesapeake Bay. This was attributed to the biomagnification of Kepone through the food chain and/or direct uptake from the water (Schimmel and Wilson, 1977). Bahner et al. (1977) confirmed this belief in a study in which spot were fed live mysids which had grazed on Kepone laden brine shrimp. Consequently, the spot accumulated concentrations of Kepone near that in their diet. Spot which had been exposed to Kepone in water were able to reduce Kepone residues in their tissues to 30-50 percent within 24-28 days in Kepone free water.

MATERIALS AND METHODS

On November 11, 1977, approximately 550 spot were obtained from the lower James River with a 30 foot semi-balloon trawl. They ranged in size from 86 mm in fork length and 8 grams in total weight to 233 mm in fork length and 165 grams in total weight. They were transported to VIMS and distributed randomly to four circular four-foot tanks (approximately 200 gallons each). All tanks were supplied with Kepone-free York River water in a flowthrough system and strong aeration. Fish were fed chopped Keponefree squid daily (8-12 percent body weight).

After one month of acclimation at ambient river temperature, three of the tanks were heated with water from a large header

tank equipped with two 220-volt heaters. Heated and unheated water were combined in mixing boxes, and flow rates were adjusted so that temperatures were maintained at approximately 22°, 17° and 12°, respectively, in the three experimental tanks. The fourth tank remained at ambient temperature except for a period between January and March in which a small heater was added to keep the water above 5°C. All tanks were insulated with cotton padding and aluminium foil. At times, temperatures in the heated tanks fluctuated as a result of sand clogging the pipes and disrupting the established flow rates. In the spring, river water temperature rose until, in June, all tanks were above 22°C. Throughout the experiment, salinity and dissolved oxygen were measured weekly, while temperatures were taken daily. In addition, water samples were analyzed periodically for Kepone.

During the acclimation period, two samples of twenty fish (five per tank) were sacrificed on Day 0 and Day 31 and analyzed for Kepone. Thereafter, biweekly samples of ten spot per tank were taken for several weeks. Since, it appeared that the spot were depurating slowly, the time interval was increased later to four weeks. Kepone concentrations (whole body, micrograms/gram, μ g/g; and parts per million, ppm) were determined by electron capture gas chromatography. Mass spectrometry was utilized when concentrations were high. For the exact methodology of the chemical analysis see Appendix A.

RESULTS

Contaminated spot depurated considerable amounts of Kepone within a period of two hundred days (Fig. 1, dotted line). A mean Kepone concentration of 1.63 ppm (N = 20) was found for spot sacrificed on the day of collection (t = 0); whereas, a mean Kepone concentration of 0.45 ppm (N = 30) was found for spot sacrificed two hundred days later (t = 200). In a statistical analysis utilizing mean concentration values for the periods t = 0 and t = 31, spot eliminated approximately 53 percent of the Kepone residues in their tissues; however, 95 percent confidence intervals were broad during this period of acclimation (Fig. 1). Bahner et al. (1977) reported residue declines of 30-50 percent in spot after 24-28 days in Kepone-free water.

Further demonstration of Kepone depuration in the spot was provided by Pearson correlation coefficients (r) of -0.7252 (p = .001) for the variables Kepone concentration with total number of days in tank (t) and -0.6231 (p = .001) for the variables Kepone concentration with total number of days in tank squared (t²). A multiple regression analysis of mean Kepone concentrations by t of each tank produced the following regression equation: Kepone concentration + 1.48183-0.145133 t + $4.5612 \times 10^{-5} t^2$, $r^2 = 0.6948$, p = .001 and the regression curve (solid line) of figure 1.

The levels of Kepone concentrations in spot varied by period (t) and by tank (Appendix B). The appearance of a rise in concentrations after day 31, when heat was applied to tanks 1, 2 and 3, was attributed to no net loss of Kepone while spot were reacclimating to the rise in temperatures (Appendix C) and to possible random samples of highly contaminated fish. Thus, the actual acclimation period for the spot might be considered as the first sixty or seventy days. Once the tanks had achieved their respective temperatures (between days 59 and 73) mean Kepone concentrations in the spot samples began to change. Spot in the warmer tanks demonstrated lower mean Kepone concentrations. In fact, spot in Tank 1 (22°C) generally exhibited lower concentrations (Fig. 1). Unfortunately, Tank 1 was discontinued after a short period of excessively high temperatures which caused a high mortality among the spot.

No significant relationships were found between the level of Kepone residues (ppm) in spot and the length, weight, or sex of the fish. Values of micrograms of Kepone per gram fish (Appendix C) produced comparable results in statistical analyses. Furthermore, no substantial growth was observed in the spot during the study period. Thus, dilution of Kepone residue in the tissues due to growth was not a factor in the rate of depuration.

Although spot and Atlantic croaker are closely related species, Kepone concentrations in spot were generally higher than those Doyle et al. (1978) found in Atlantic croaker. Both species were collected from the James River at approximately the same time of year (October-November) although in different years (1976-1977). Initially, spot depurated Kepone at a faster rate than Atlantic croaker (Fig. 2). In fact, there was no significant decrease in Kepone levels of Atlantic croaker until after a period of fiftysix days. On the other hand, Atlantic croaker that were sacrificed after a period of one hundred and fourteen days, depurated at a slightly faster rate than spot from Tank 4 (ambient temperature).

CONCLUSIONS

Spot, like other fishes, depurated Kepone at a slower rate than some invertebrate species (see: Bahner et al. 1977). A mean loss in Kepone residues of 72 percent occurred between the initial spot sample (t = 0) and the eight spot sample (t = 200). A plot of the variables, mean Kepone concentration by period (t) (Figure 1) demonstrated the fact that a negative relationship existed between Kepone concentration in spot and the amount of time a spot was allowed to depurate in Kepone free water. Nonetheless, only 30 percent of the spot (N = 309) utilized in the test were below the established action level for human consumption (0.3 Therefore, it appears that it would be impractical to re-. (mag move spot from a contaminated area and to maintain them in a holding facility for the purpose of depuration and later commercial sale. Whether wild spot from the James River and the lower Chesapeake Bay can or cannot eliminate Kepone from their bodies while in the overwintering grounds of Virginia and North Carolina is still another question. To answer this question and other management questions, we would have to establish the Kepone levels in fish from offshore and returning populations which would be very difficult and costly.

Temperature was an important factor in the rate of Kepone depuration in spot. Spot held in warmer water exhibited lower mean Kepone concentrations; however, we were unable to observe the effect of the lower temperature extremities for any length of time in the cooler tanks because of the rise in temperature during the later spring months. In response to the warmer temperatures, Kepone concentrations in spot indicated that the rate of elimination of Kepone from body tissues is probably a function of the rate of an individual's metabolism. Thus, an increase in the matabolic rate as a result of an increase in body temperature may cause an acceleration in the depuration rate; however, it may not be apparent until after a period of acclimation.

It is regretable that the cost of Kepone body burden analysis is so high that sample sizes must remain small. In the future, we should take a closer look at the processes of uptake and accumulation of Kepone in eggs, larvae, juvenile and adult life stages. Also, we must have a better understanding of how Kepone concentrations in fish are related to uptake, accumulation and the lipid composition of fish.

- Bahner, L. H., A. J. Wilson, Jr., J. M. Sheppard, J. M. Patrick, Jr., L. R. Goodman, and G. E. Walsh. 1977. Kepone Bioconcentration, Accumulation, Loss and Transfer through Estuarine Food Chains. Chesapeake Science. 18:299-308.
- Bender, M. E., R. J. Huggett, and W. J. Hargis. 1977. Kepone Residues in Chesapeake Bay Biota. Kepone Seminar II, September 20 and 31, Easton, Maryland. MMS available from author.
- Doyle, R. T., J. V. Merriner, and M. E. Bender. 1978. (In Press) Depuration of Kepone by Atlantic Croaker (<u>Micropogonias</u> <u>undulatus</u>) in a Laboratory Study. Proc. 32nd Ann. Conf. SE Fish and Game Comm.
- Schimmel, S. C. and A. J. Wilson, Jr. 1977. Acute Toxicity of Kepone to Four Estuarine Animals. Chesapeake Science. 18:224-227.

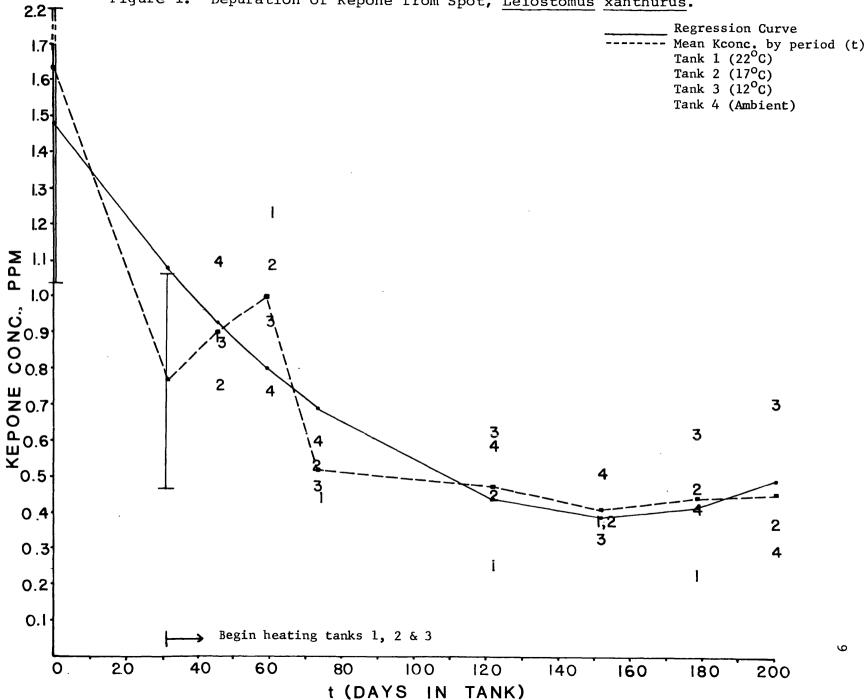
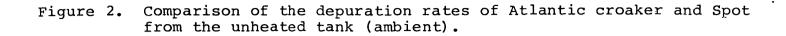
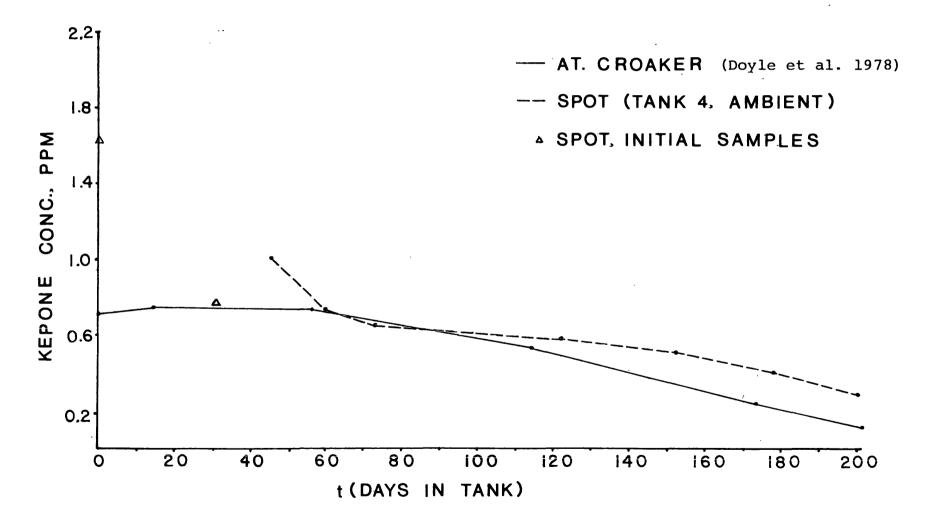


Figure 1. Depuration of Kepone from Spot, Leiostomus xanthurus.





Appendix A

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Chemical Analysis for the Pesticide Kepone

Appendix A

Chemical Analysis for the Pesticide Kepone

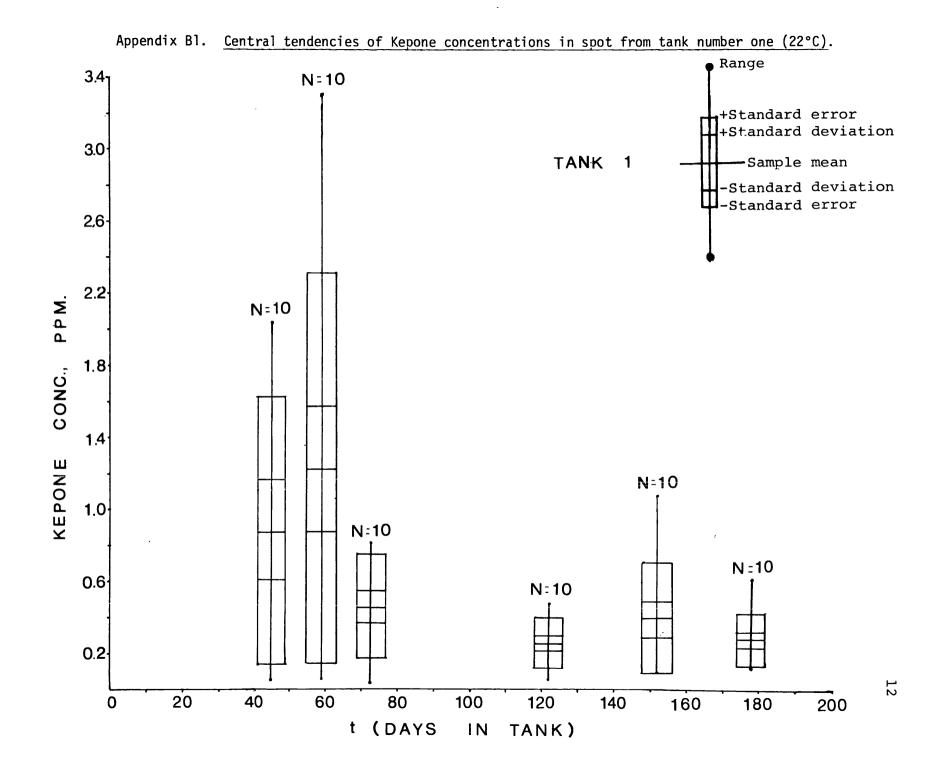
Whole fish were ground in a meat grinder into hamburger consistency. A mixture of anhydrous sodium sulfate and Quso^R G-30 (precipitated silica, Philadelphia Quartz Co.) was added for desiccation. The proportions of sample to the desiccants 30 g fish - 54 g Na, SO₄ - 6 g Quso. Then samples were were: frozen at -5°C for 24 hours to rupture the cells. After thawing the desiccated samples were ground with a blender to a powdery consistency and transferred to pre-extracted paper thimbles for Soxhlet extraction. Extraction was carried out using 1:1 ethyl ether-petroleum ether for 16 hrs. Extracts were then concentrated by evaporation and cleaned by activated fluorisil column chromatography (EPA, 1975). The Kepone containing elutriate was analyzed by electron capture gas chromatography utilizing packed columns with one or more of the following liquid phases: 4% SE-30 + 6% OV 210; 1.5% OV-17 + 1.95% QF-1 + 3% CV-1. On occasion, when concentrations were sufficiently high, Kepone presence was confirmed by mass spectrometry.

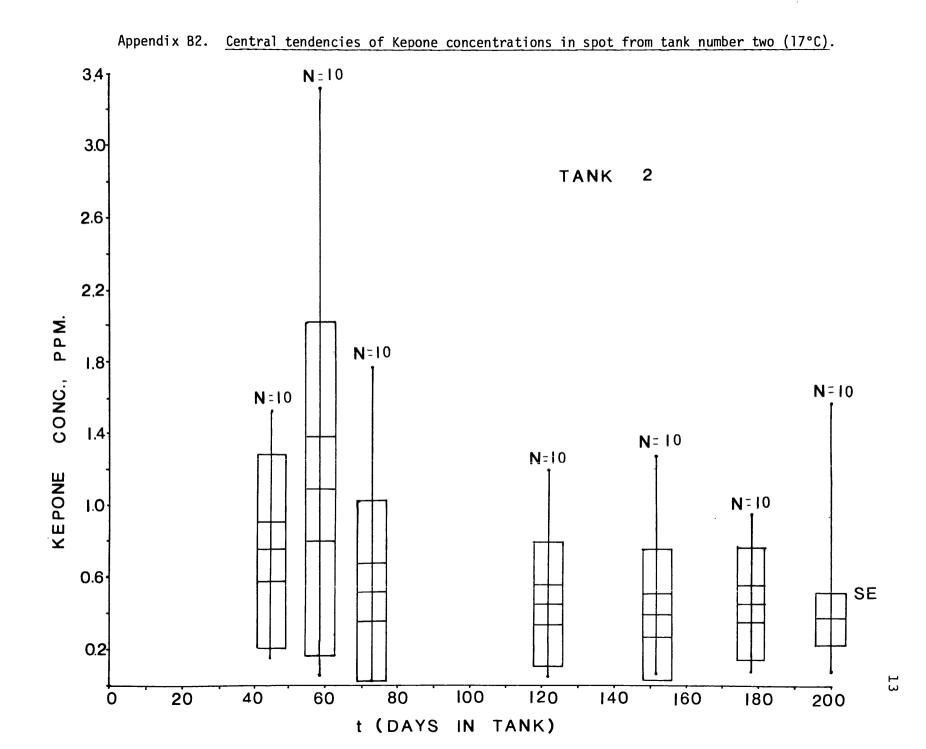
Appendix B

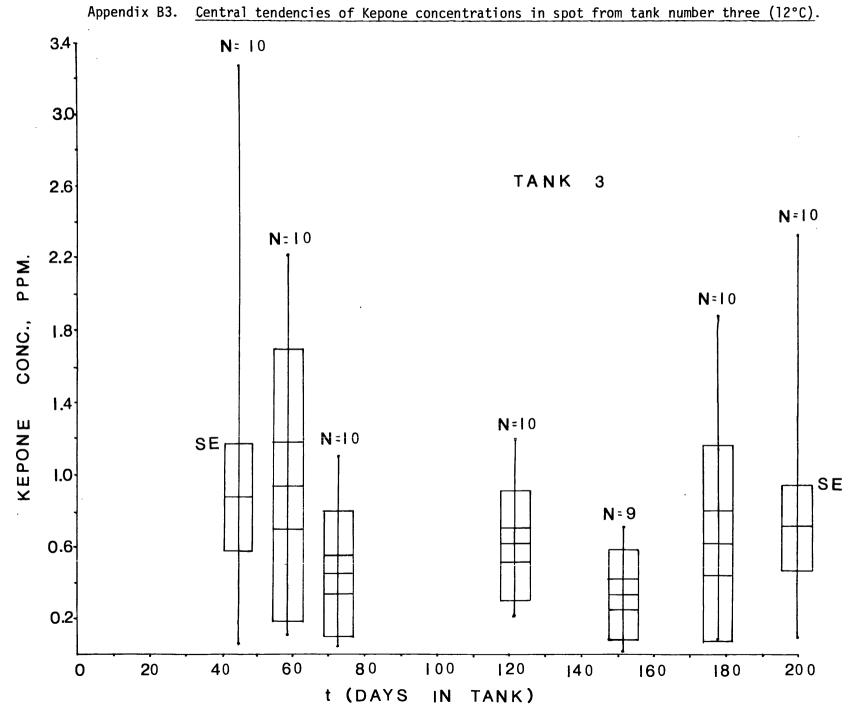
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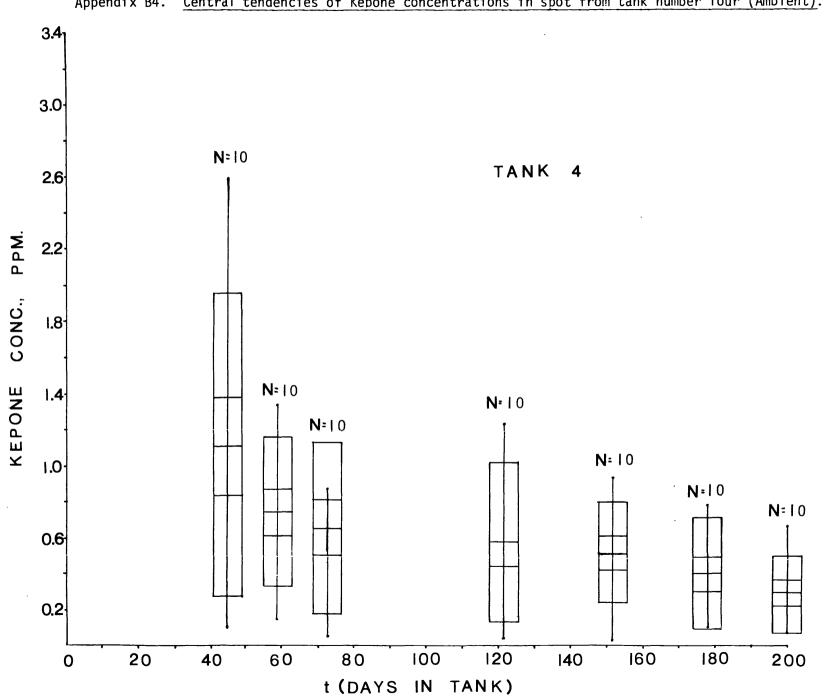
Descriptive Statistics of Kepone Concentrations Broken Down by Tank and Sampling Period

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REGRESSICN ANALYSIS FER SPCT DEFURATION STUDY			Appendix B5.	11/27/78		3		
ILE CSPCT	(CREATICN CATE = 1		ams of Kepone per gram 1					
RITERICN VARIAN ERCKEN COWN	BLE MGK EV TCAY TO	CESCRIPTIG TAL NUMBER CF DAYS IN CICATES ONE CF FGUR CG	TANK	PULAIIUN	· · · · · · ·			
							•	
FARIABLE	CGEE	VALLE LABEL	SUM	MEAN	STD DEV	VARIANCE		N
FOR ENTIRE POPUL	LATION		7539.5654	24.3212	38-5215	1483-9021	(310
TCAY	0.	FRESAMPLE	686.1836	34. 30 92	79.9743	6395.8877	(20
TANKAC	0.		686.1836	34.3092	79.5743	6395.8877	(20
TEAY TANKNC	31. C.	IST SAMPLE	516.9828 5 76. 9828	28.8491 28.8491	39.3826 35.3826	1550.9912 1550.9512	(.	. 20 20
ΙΓΑΥ	45.	SECCNE SAMPLE	1178.6245	29.4656	37.4720	1404.1498	(-	40
TANKAC	1.		247.6199	24.7620	27.4161	751.7521	1	10
TANKAC	ž.		178.1039	17-8164	19.4443	378.0817	ť	10
TANKNC	3.		331-1999	33.12CC	50.6346	2563.8608	(10
1 ANK NC	4.		421.7608	42.1701	44.9584	2021-2588	ſ	10
1C#Y	55.	THIRC SAMPLE	1628-8995	40.7225	53.5146	2663-8095	(4(
TANKNC	1.		464.3159	46.4320	67.9159 53.9748	4612.5746 2913.2750	l	10
TANFNC	2.		431.5599 337.5699	43.156C 33.757G	46.7663	2378.1477	ł	10
TANKNC TANKNC	3 • 4 •		395.4498	39.5450	49.0672	2407.5916	ſ	10
CAY	73.	FCURTH SAMPLE	830.1347	20.7534	27.4251	752.1352	(40
TANKAC	1.		128.2000	12.8200	£.C695	65.1168	(10
TANKAC	2.		211.1755	21-1180	36.1400	908.4175	i i	10
TANKNC	3.		185.8059	18.5810	15.2741	371.4914	(10
TANKAC	4.		304.5449	30-4545	41.6350	1733-4711	l	10
TEAY	122.	FIFTH SAMPLE	717.5548	17.9389	21.5632	464.9730	ł	40
TANKNC	1.		71.1500	7.115C	6.7327 19.6316	45.3292 385.3599	1	10
TANKNC	2.		167.65CC 305.4645	16.7850 30.9465	30.0739	504.4377	i	10
1ANKNC TAN¥NC	3. 4.		165.0859	16.9090	18.5443	358.8867	i	10
TCAY	152.	SIXTE SAMPLE	651.4358	16.2860	22.4033	501.5059	(40
TANKNE	1.		150.5900	15.0590	17-1141	292.8920	(10
TANKNE	ž.		242.6799	24.2680	36.3176	1466.2410	(10
TANKNC	3.		65.1SCC	€.5150	5.4601	29.8123	1	10
TANKAC	4.		152.5755	15-2980	13.9801	195.4421	(10
TCAY	178.	SEVENTH SAPPLE	736.5857	18.4147	24.4548	559.9951	(40
TANKNC	1-		85.3500	8.9350	11.0222	121.4899	({	10
TANKNC	2.		147.53CC	14-7530	11.2755	127.1375	i	10
TANKNE	3.		375.0198	37.5020	41.4401	1717.2845	(10
TANKNC	4.		124.6900	12.4690	8.6738	75.2349	(10
TEAY	200-	FINAL SAPPLE	533-1599	17.7720	31.6315	د2 1000،552	t	30
TANKNC	2.		136.4200	13.0420	16.0274	256-0764	(10
TANKNC TANKNC	3.		253-8000	29.3800	51.1105	2612.2806	ŧ	10
CENERCE.	4.		108.9400	10.8940	11.3007	127.7056	í	10

REGRESSION ANALYSIS FOR SPOT DEPURATION STUDY			A 11 - F4	11/27/78 PAGE 7				
		(27/78) Mean length in	Appendix B6.	ot camples				
FILE ESFET (CR	EATILN LATE = 1.	Mean rength m	minimeters (mm) or sp	ot samples.				
		CESCRIPTICA	CF SUEPO	PULATION	s		-	
CRITERICN VARIABLE		IAL LENGTH IN MILLIMETE			•			
ERCKEN COWN BY		TAL NUMBER OF DAYS IN T		-				
EY	TANKNE IN	LICATES ONE OF FOLR CON	TRCL TEMPERATUR					
VARIABLE	COLE	VALLE LABEL	SUP	MEAN	STD DEV	VARIANCE		
		VALLE LADEL				•		
FCR ENTIRE POPULATI	CN		401C5.000C	125.3710	25.3308	641-6516		
TEAY	ί.	FRESAMPLE	1945.CC00	97.2500	11.1255	123.7763		
TANKAC	C.	TPECAPTEE	1945.0000	97.2500	11.1255	123.7763		
	21	IST SAMFLE	2599.0000	129.9500	33.3868	1114.6816		
TC/Y TANKNC	31. C.	IST SAFFLE	2595.0000	129.9500	33.3868	1114-6816		
TANK TO BE								
TCAY	45.	SECCNE SAPPLE	4505-0000	122.7250	16.4986	272.2045		
TANKNC	1.		1154-COCO 1143-COGO	115.4COC 114.3COC	10.5325 12.7545	11C.9333 162.6778		
TANKNC Tanknc	2. 3.		1279.0000	127.5000	18.5379	343.6556		
TANKAC	4.		1293.0000	129.3000	19.8329	393-3444		
		TI 105 CANDI 6	6:21 0000	130-5250	25.1630	633.1788		
TANKNE	59 . 1-	THIRE SAMPLE	5221.COCO 1273.COCO	127.3000	20.1332	405.3444		
TANKNC	2.		1279.0000	127.9000	26.7266	714.3222		
JANKAC	3.		1272.CCGC	127.2000	21.7552	473.2889		
TANKAC	4.		1397.0000	139.7000	32.0275	1025.7869		
TEAY	73.	FCURTH SAMPLE	5441.COGC	136.0250	26.2068	686.7942		
TANKAC	1.		1314-060C	131.4000	21.9555	482.0444		
TANKAC	ź.		1398.CCCC	139.8000	26.8816	722.6222		
TANKNE	3.		1358-0000	135.8000	25.4157	645.5556		
ΤΑΝΚΝΟ	4.		1371-0000	137.1000	32.9324	1084.5444		
TCAY	122.	FIFTH SAMFLE	5314.0000	132.8500	25.0983	625.5256		
TAVKAC	1.	•	1265.0000	126.9000	21.0895	444.7667		
TANKNC	2.		1361.0000	136.1000 141.4000	3C.2120 26.7362	912.7667 714.9333		
TANKAC	3. 4.		1414.000C 127C.CCCC	127.000	22.0656	466.8889		
122400	7.		12 10 0000					
TDAY	152.	SIXTH SAMPLE	5365.0000	134.1250	27.2333	741.6506		
TANKNC	1-		1295.0000	129.5000 144.8600	28.2813 30.1065	755.8333 966.4660		
TANKAC	2.		1448.COCO 1255.CCCC	125.9000	26.4801	419.4333		
TANKNE	3.		1363.CCCC	136.3000	25.2804	857.3444		
JANKNC	4-		12020000	1301 3000				
TDFA	173.	SEVENTH SAPPLE	534C.COCO	133.5COC 120.7GOG	21.5585 16.8518	466.5128 358.5000		
TANKNC	1.	-	12C7-COCC 1358-COCO	135.6000	21.6220	467.5111		
TANKNC	2.	•		13360000	61466CV	10102111		
T &NKNC T ANFNC	3.		1477.0000	147.7000	24.9491	622.4556		
IMNENL	. 4.		1298-0000	129-8000	12.1454	147.5111		
TEAY	200-	FINAL SAMFLE	3571.0000	132.3667	22.2734	496-1023		
TANENC	2.		1332-0000	133.2000	23.4132	548.1778		
TANFNE TANKNE	3.		1301-000	130.1000	21.4965	462.1000		
145651	4.		1338.0000	133.8000	24-0730	579.5111		

TCTAL CASES = 310

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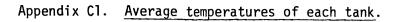
	LYSIS FCF SPCT DEPURA		Appendix B7.	11/27/7	8 FAGE	FAGE 11		
ILE DSPCT		.127/76) <u>Mean weight i</u>						
RITERICN VARIA BRCKEN DOWN	ABLE WEIGHT TOT	CESCRIPTIG ALBEIGHTINGRAPS ALNUPBERGFCAYSIN		PULATION				
		ICATES CNE GF FCUR CO						
VAFIABLE	CCCE	VALUE LABEL	SUM	MEAN	STD CEV	VARIANCE		ı
FCF ENTIPE FUP	ULATION		16561.8995	35.1674	25.8903	670.3069	ť	310
ICAY	с.	PRESAMPLE	320.6000	16-030C	15.2458	370.5548	t	2
TANKNC	0.	:	320.6000	16.0300	15.2498	370.5548	ł	20
TANKNC	31. C.	IST SAMFLE	678.0C0C 678.CCCC	33.9CCC 33.9CCC	32.7316	1071.3575 1071.3579	(t	20
							۰.	
CAY	45.	SECCNC SAMPLE	1126.3000	28.1575	13.9141	193-6020	(4
TANKNE	1.		235.0000	23.9000	7.8521	61.6556	(1
TANENC	2.		215.7000	21.9700	9.C62C 14.5194	82.1201 210.6135	l	1
TANKNC TANKNC	. ژ 4.		32C.70CG 346.900C	32.070C 34.650C	16.5011	357.2495	Ċ	1
6 41			1416 6666	35.3750	27.0135	724.1276	(4
ICAY	55.	THIRC SAMPLE	1415.CCCC 285.CCCC	26.5000	15.4722	235.3685	i i	1
ΊΔΝΚΝር ΊΔΝΚΝΟ	1. 2.		231.0000	33-1000	23.0287	530.3222	ì	i
TANKNE	3-		333.GOCU	33.3000	23.1855	537.5667	i	1
TANENC	4.		466.0000	46.6000	46.6426	1651.8222	i	10
TEAY	73.	FCURTH SAPPLE	1556.0000	36.9006	25.8601	666.7462	C	4
TANKNC	1.		328.0000	32.8000	16.6986	£70=8444	(10
TANKAC	2.		416.0GUC	41.6000	22.2221	453.8222	(1
TANKIC	3.		386.5000	36.0500	25.7601	663.5206	(10
TANKNC	4.		431.5000	43.1500	37.3006	1351-3361	(10
1641	122.	FIFTH SAMPLE	1443.0000	36.C75G	26.C713	675.7122	(4
TANKNC	1.		282.0000	28.2000	21.2697	452.4060	(1.
TANKNE	2 .		428.0000	42.8000	37.5227	1407.4556	(10
TANKNE	3.		430.0000	43.0000	23.4852	551.5555	l L	10
TANKNC	4.		3C3.CC00	30.3000	18.0003	324.0111	•	10
TCAY	152.	SIXTE SAMPLE	1557.CCCC	35.9250	29.8641	891.8660	(4
TANKNC	1.		375.0000	37.5000	31.6763	1003.3855	(1
TANKNC	ž.		496.0000	45.0000	25.0593	844.4444	(1
TANKNO	3.		260.0000	28.0000	15.2607	232-8585	(1
TANKNC	4.		452.0000	45.200C	38.6919	1457.066?	,	10
TCAY	176.	SEVENTH SAMPLE	1535.0000	38-3750	23.5260	553.4712	ſ	4
TANKAC	1-		262.0000	26.2000	14.7463	217.5111	(1
TANKNC	ž.,		4C3-CCGC	46.3000	30.1835	953.7889	(1
TANKNC Tanknc	3.		517.0000	51.7000	26.8537	721.1222		21
	4.		353.0000	35.3000	11.3730	129.3444	C	ì
TCAY	200.	FINAL SAMPLE	1231.0000	41.0333	27.8524	775.7575	ι	3
TANKNO	2.		437.0000	43.7000	32.4004	1645.7889	(1 (
TANKNE	3.		351.000	35-1000	18.5270	335.8710	(10
TANKNE	4•		443.GOCC	44.3000	32.4826	1055.1222	(10

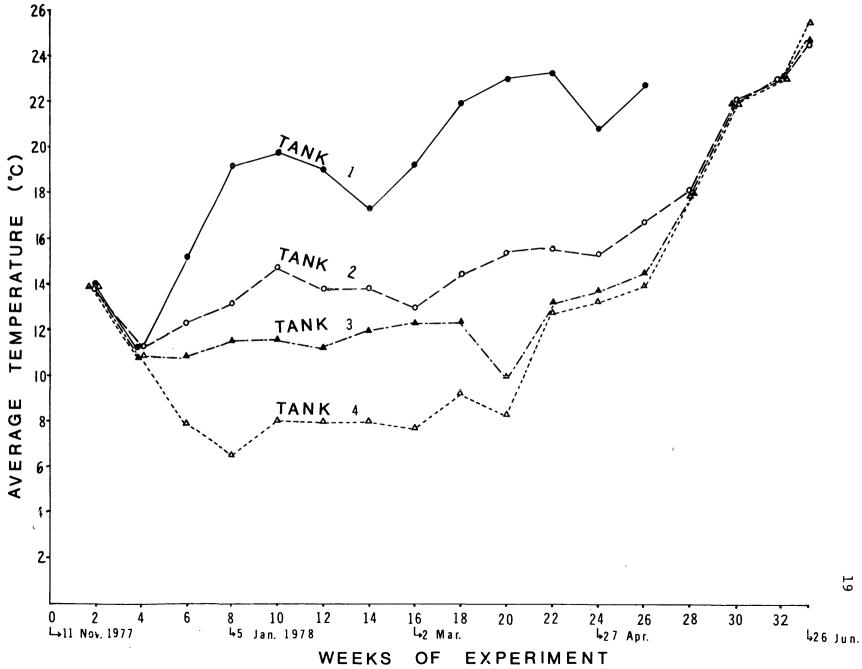
1CTAL CASES = 310

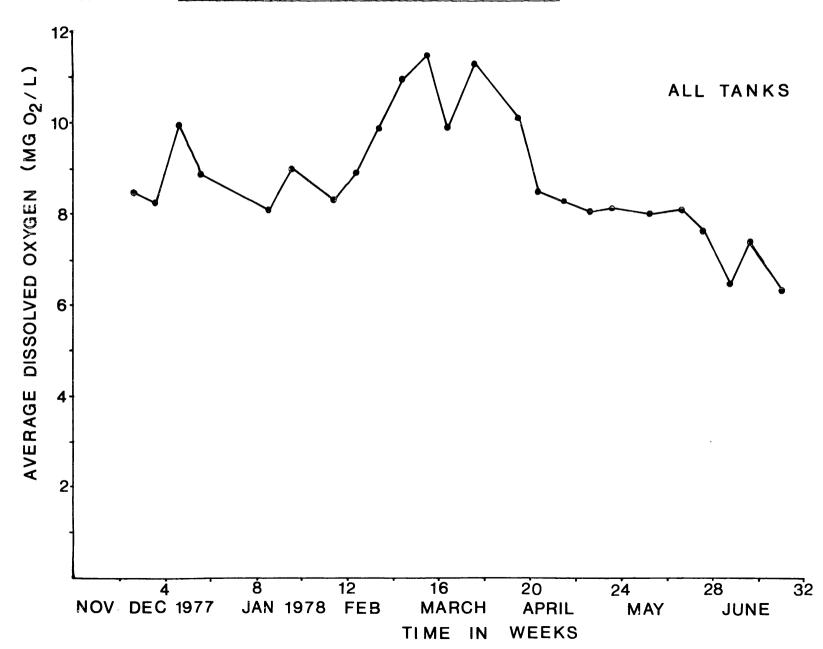
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Appendix C

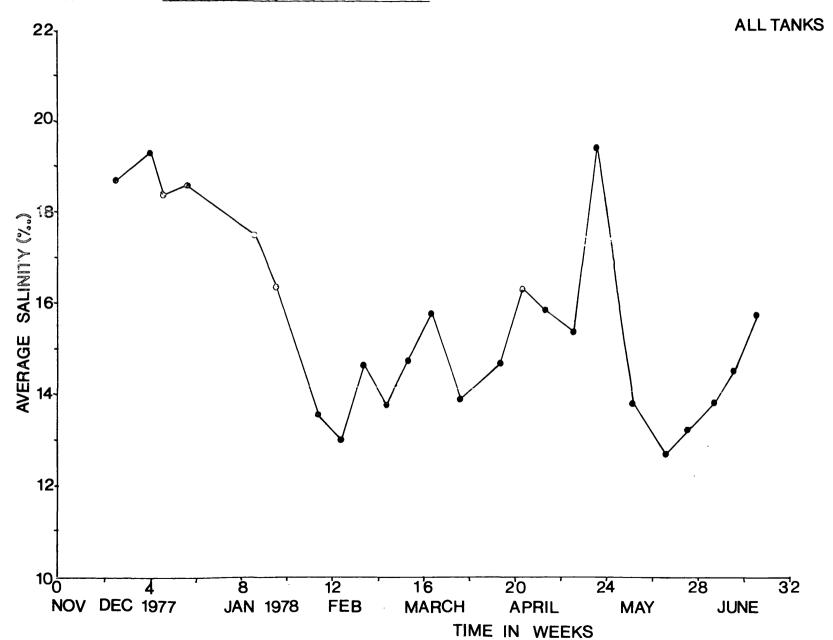
Water Quality Analysis During the Experimental Period







Appendix C2. Dissolved oxygen concentrations for all four tanks.



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Appendix C3. Average salinity for all four tanks.